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Qualitative and quantitative demonstration of bead-to-bead transfer with bone marrow-derived human mesenchymal stem cells on microcarriers: utilising the phenomenon to improve culture performance

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Highlights

- Qualitative & quantitative demonstration of bead-to-bead transfer for hMSC microcarrier culture
- The phenomenon is used to significantly improve yield and culture performance
- Reproducible results across 3 different hBM-MSD donors in FBS, HPL and serum-free

Abstract

Human mesenchymal stem cells (hMSCs) are a key candidate for advanced cell therapies with numerous clinical trials investigating their potential to treat acute and chronic indications. However, important translational and manufacturing challenges need to be addressed to improve our capability for scalable production of fully functional cells. In this study, we have demonstrated, both qualitatively and quantitatively, the ability of bone marrow-derived hMSCs to migrate from one microcarrier to another, and, to populate fresh microcarriers when added into suspension culture. Additionally, we have shown that compared to inoculating a culture with cells in free suspension, inoculating 10% of near-confluent microcarriers from an initial seed microcarrier culture resulted in an increase in the cell growth rate and overall cell yield and a significant reduction in the lag phase. These findings were consistent across cells from three different BM-hMSC donors and across different culture medium conditions, foetal bovine serum-supplemented medium, human platelet lysate-supplemented medium and serum-free medium. This new cells-on-beads inoculation method is an effective means of process intensification with the potential to decrease manufacturing times and potentially costs of hMSC-based therapies.

Keywords: human mesenchymal stem cells; cell therapy bioprocessing; microcarriers; bioreactor; regenerative medicine; bead to bead transfer; process intensification

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1. Introduction

Advanced therapies, including cell and gene-based therapies, will form the next generation of therapeutics which have the potential to address unmet clinical need, treat chronic conditions and offer potential cures rather than simply alleviating patient symptoms (Barry and Murphy 2013). Adherent, or anchorage-dependent cells, are likely to be promising candidates for both autologous (patient-specific) and allogeneic (universal donor) cell-based therapies; such cells include human adult mesenchymal stem cells (hMSCs), human embryonic stem cells (hESCs) and human induced pluripotent stem cells (hiPSCs). However, the growth of adherent cells presents additional challenges in comparison to suspension cell platforms, and has therefore been the subject of considerable research activity (Rafiq 2016). Initially in the 1980s and 1990s, significant effort was directed at the production of adherent Chinese Hamster Ovary (CHO) cells for protein/vaccine production, and more recently, for the production of adherent stem cells for cell therapy applications. Established techniques for the growth of these cells involve monolayer culture, predominantly in tissue-culture flasks or roller bottles. However, as was recognised for adherent CHO cells in the 1980s, alternative manufacturing processes for the production of adherent cells are required which are scalable, robust, reproducible and cost-effective. This is achieved through the use of microcarriers in stirred-tank bioreactor systems and have been used for multiple cell types including CHO (Ohlson *et al.* 1994), hMSCs (Dos Santos *et al.* 2011; Rafiq *et al.* 2013a), hESCs (Chen *et al.* 2010) and hiPSCs (Badenes *et al.* 2017; Bardy *et al.* 2013).

In monolayer culture, cell growth is maintained by keeping the cells in the log phase of the culture and is based upon 'confluency', the proportion of adherent cells covering the attachment surface. When cells reach an appropriate confluency level (usually 80% but can differ based on cell type), the cells are removed from the surface and plated into new tissue culture flasks in order to keep them in the exponential phase, a process known as passaging. The process of passaging typically

requires the use of a proteolytic enzyme such as trypsin, dispase or collagenase (Lindner *et al.* 1987) in conjunction with a chelating agent such as EDTA. The exposure of adherent cells to these reagents in the absence of serum-containing medium enables the digestion and cleaving of cell attachment proteins. However, frequent exposure to such reagents has been demonstrated to have adverse effects on mammalian cells including proteome alteration (Huang *et al.* 2010) and effects on cell shape and chromatin structure (Kapiszewska *et al.* 1991). As such there is concern regarding the consecutive passaging of cells, in part due to the long-term deleterious effects of dissociation reagents on the cell cytoskeleton (Madeira *et al.* 2012). Although enzyme-free dissociation buffers also exist, some formulations may not be suitable for use with hMSCs. For example, a study using a Gibco-brand enzyme-free dissociation buffer to detach hMSCs grown on tissue culture plastic found that not only was cell viability following detachment significantly reduced compared to trypsin controls (~ 70% compared to > 90%, respectively) but cells also failed to reattach to tissue culture plastic (Heng *et al.* 2009).

Until now, hMSC microcarrier culture has followed the same approach as monolayer culture with respect to harvesting when cells reach confluency, *i.e.* the use of enzymes to detach cells from the surface of the microcarriers, preferably aided by agitation to reduce the time the cells are exposed to that environment (Nienow *et al.* 2014). However, unlike monolayer culture which is limited by the surface area available in the tissue culture flask, microcarrier/bioreactor culture has a significant advantage with respect to surface area to volume ratio and there is potential for the provision of additional microcarriers during a culture process when the cells are nearing confluency in order to maintain cell growth in the exponential phase for longer. For example, Hervy and colleagues recently maintained hMSCs on microcarriers for over 40 days using this approach (Hervy *et al.* 2014). This extended culture time relies on the transfer of cells from one microcarrier/bead to another (called here for convenience, bead-to-bead transfer). Bead-to-bead transfer (or cell

migration from one microcarrier to another) during suspension culture of adherent cells is believed to be a key mechanism for rapid and efficient cell expansion in suspension cultures (de Soure et al. 2016). This cell behaviour is of particular interest from a process development and manufacturing perspective as the demonstration of bead-to-bead transfer would facilitate the large-scale manufacture of stem cells via seed bioreactor cultures without the need to expose the cells to frequent deleterious proteolytic enzymes. This approach also facilitates process intensification by providing additional surface throughout the course of the culture, maintaining the cells in the exponential phase of growth for a longer period of time, thereby increasing yield and efficiency whilst reducing process costs and time.

Although investigated and proven for adherent CHO and Vero cells (Ohlson et al. 1994; Wang and Ouyang 1999), work demonstrating such a phenomenon for hMSCs has to date focused on single hMSC donors (Takahashi et al. 2017), or an immortalised cell line (hMSC-TERT) (Leber et al. 2017) or it included complex agitation regimes (Hervy et al. 2014). In this study, we demonstrate, both qualitatively and quantitatively, the phenomenon of bead-to-bead transfer for three donor BM-hMSCs on different microcarriers (Plastic P102-L, Cytodex-3 and Hillex) in different culture medium conditions, foetal bovine serum (FBS), human platelet lysate (HPL) and serum-free medium (SFM). We also explore the use of a cells-on-beads inoculation procedure, which subsequently utilises bead-to-bead transfer (henceforth called 'cells-on-beads inoculation'), in contrast to the standard free suspension inoculation procedure to facilitate process intensification. Several important parameters were measured throughout the culture including cell growth kinetics, viability, key metabolite consumption and production, immune-phenotype and differentiation potential.

2. Materials and methods

2.1 hMSC monolayer expansion

Human MSCs from three donors were used in this study. All were isolated from bone-marrow aspirate and obtained from Lonza (Walkersville, USA) after the donors provided informed consent. The university's local Ethical Committee approved the use of the samples for research. The isolation and culture of primary hMSCs is described in Rafiq *et al.* (2013b); in brief, hMSCs were isolated on the basis of plastic adherence and cryopreserved at passage 1 at a density of 2×10^6 cells/mL in 10 % dimethyl sulphoxide (DMSO) (v/v) (Sigma Aldrich, UK) and 90 % foetal bovine serum (FBS; Hyclone, Lot# RUF35869). The cells were cultured according to previously determined protocols (Heathman *et al.* 2016; Heathman *et al.* 2015a; Heathman *et al.* 2015b; Rafiq *et al.* 2013a). Briefly, the cells were seeded at 5,000 cells/cm² and grown on tissue culture plastic in DMEM (1 g/L glucose, Lonza, UK) supplemented with 10% (v/v) foetal bovine serum (FBS; HyClone) or 10% (v/v) non-heparin requiring (PL-NH) Stemulate™ (Cook Regentec, USA) for the human platelet lysate based culture conditions, and 2 mM UltraGlutamine (Lonza, UK).

Where cells were also cultured under serum-free medium (SFM) conditions, the Prime-XV™ SFM hMSC medium was used (Irvine Scientific, USA) in accordance with the manufacturer's instructions. As required, attachment surfaces were pre-coated with recombinant fibronectin (Irvine Scientific, USA). For both the SFM and HPL cultures, the BM-hMSCs underwent one adaptation passage in medium containing either the Prime-XV™ medium or 10% (v/v) PL-NH Stemulate™.

2.2 Spinner flask microcarrier culture

The spinner flask cultures involved the use of 100 mL BellCo spinner flasks (BellCo, USA), with a 100 mL working volume and a vessel diameter (T) of 60 mm. The spinner flask vessels were siliconized prior to use with Sigmacote (Sigma Aldrich, UK) as described in (Rafiq *et al.* 2016). The spinner flasks were agitated by a magnetic stirrer bar at the just suspended speed (N_{JS}) placed on the Bell-Ennum™ Compact 5 position magnetic stirrer platform (BellCo, USA) with air in the

headspace for surface aeration giving essentially 100% dO₂. The microcarriers were prepared according to the manufacturer's instructions and sterilised by autoclaving at 121°C and 1 atm for 15 minutes. Prior to use, the microcarriers were conditioned in growth medium for at least 1 hour to facilitate cell attachment. The surface area provided per spinner flask was 500 cm² and the cells were inoculated at a density of 6,000 cells/cm² as previously described (Rafiq *et. al.*, 2013; Heathman *et. al.*, 2015). This inoculation method, referred to later as the 'control', is in contrast to the 'cells-on-beads' inoculation method (discussed later in the results section) which involved inoculating 10% of near-confluent microcarriers from an initial seed microcarrier culture. In all cases, a 50% medium exchange was performed at 72 h in culture and then every 48 h thereafter.

2.3 Analytical Techniques

Cell counts and viability (via acridine orange uptake and DAPI exclusion) were determined using the Nucleocounter NC-3000 (Chemometec, UK). Cell viability was also assessed by the Live/Dead Viability/Cytotoxicity kit (Calcein AM/ Ethidium Homodimer; Life Sciences, ThermoFisher, UK) following the manufacturer's instructions. Briefly, the cells were first washed with PBS and then incubated with the Live/Dead stain containing a final concentration of 2 μM calcein-AM and 4 μM ethidium homodimer in the dark at 37°C and 5% CO₂ for 40 minutes. Post incubation, the Live/Dead working solution was removed and replaced with PBS. Samples were then visualised on a Nikon Ti Eclipse epi-fluorescence microscope (Nikon, UK).

Analysis of glucose and lactate concentrations in the spent medium was performed using a Cedex Bio-HT (Roche, Germany). The following parameters were calculated from the data obtained:

1. Specific Growth Rate

$$\text{Specific growth rate, } \mu = \frac{\ln\left(\frac{C_x(t)}{C_x(0)}\right)}{\Delta t}$$

where μ is the net specific growth rate (h^{-1}), $C_x(t)$ and $C_x(0)$ are the cell numbers at the end and start of the exponential growth phase, respectively and t is time (h).

2. Population Doublings

$$\text{Population Doublings, } P_d = \frac{1}{\log(2)} \cdot \log\left(\frac{C_x(t)}{C_x(0)}\right)$$

where P_d is the number of population doublings, $C_x(t)$ and $C_x(0)$ are the cell numbers at the end and start of the exponential growth phase, respectively.

3. Specific Metabolite Consumption/Production Rate

$$\text{Specific metabolite flux, } q_{met} = \left(\frac{\mu}{C_x(0)}\right) \cdot \left(\frac{C_{met}(t) - C_{met}(0)}{e^{\mu t} - 1}\right)$$

where q_{met} is the net specific metabolite consumption or production rate, μ is the specific growth rate (h^{-1}), $C_x(0)$ is the cell number at the end of the exponential growth phase, $C_{met}(t)$ and $C_{met}(0)$ are the metabolite concentrations at the end and start of the exponential growth phase, respectively and t is time (h).

2.4 hMSC Characterisation

Immunophenotype analysis was performed by multiparameter flow cytometry before and after the hMSC expansion process using a previously developed protocol (Chan *et al.* 2014). In brief, this involved the use of flow cytometry where harvested, single cells were suspended in growth medium and loaded onto a 96 well plate. The cells were stained with monoclonal antibodies against the positive hMSC markers, CD73, CD90 and CD105 and the negative markers, CD34 & HLA-DR (BD Biosciences, UK) in addition with the corresponding isotype controls (BD Biosciences, UK). The antibodies selected were those recommended by the International Society for Cellular Therapy (ISCT) for the characterisation of hMSCs (Dominici *et al.* 2006).

The hMSC differentiation was induced using Prime-XV™ Differentiation SFM (Irvine Scientific, USA) as per manufacturer's instructions. After 21 days, the differentiation media were removed, cells rinsed with PBS then fixed with 4% (v/v) PFA at room temperature. Adipocytes were stained with 1% (w/v) Oil Red O (Sigma-Aldrich, UK) in isopropanol at room temperature and rinsed with distilled water. Osteoblasts were incubated with 2.5% (v/v) silver nitrate (Sigma-Aldrich, UK) under ultraviolet light (30 minutes exposure), rinsed with distilled water and stained with fast violet solution (Sigma-Aldrich, UK) containing 4% (v/v) naphthol AS-MX phosphate alkaline (Sigma-Aldrich, UK) for 45 minutes at room temperature in the dark. Chondrocytes were stained with 1% (w/v) Alcian blue (Sigma-Aldrich, UK) in 0.1 M hydrochloric acid (Sigma-Aldrich, UK). After 30 minutes incubation, cells were rinsed three times with 0.1 M HCl. After staining, differentiated cells were visualized under a light microscope (Nikon Eclipse TS-100, UK).

2.5 Tracking of bead-to-bead cell transfer:

Bead-to-bead cell transfer was assessed by two methods: 1) by employing visually distinctive microcarriers in the same culture system (adding a visually distinctive microcarrier at a later stage of the culture) and 2) by fluorescently labelling cells with two different Cell Tracker™ dyes (Life Technologies, ThermoFisher Scientific, UK). The use of visually distinctive microcarriers provides an effective way of demonstrating bead-to-bead transfer. The cells were originally cultured on the Plastic P102-L microcarriers in agitated spinner flask conditions (as described above). After 3 days of culture, the visually distinctive microcarriers (either the Hillex or Cytodex-1) were added at the same microcarrier concentration during a 50% medium exchange. The culture was continuously agitated throughout the addition of the second microcarrier type so that the microcarriers were never static. This procedure unequivocally demonstrated the transfer of cells from the Plastic P102-L microcarrier to the new microcarrier (either Hillex or Cytodex-1).

For the fluorescent labelling, Cell Tracker™ Green CMFDA and Cell Tracker™ Red CM-Dil (Life Technologies, ThermoFisher Scientific, UK) were employed. Cell Tracker™ Green contains chloromethyl derivatives of fluorescein diacetate (CMFDA) and is nonfluorescent until cytosolic esterases cleave off the acetates, releasing a brightly fluorescent product. Unlike the Cell Tracker™ Green CMFDA, Cell Tracker™ Red CM-Dil is a lipophilic carbocyanine that incorporates a mildly thiol-reactive chloro-methyl substituent with affinity for the lipids present inside the cell membrane. These reagents pass freely through cell membranes, but once inside the cell, are transformed into cell-impermeant reaction products.

The Cell Tracker™ dyes were used according to the manufacturer's instructions. Briefly, based on the average cell concentration obtained post-harvest, the volume of cell suspension required for each experiment to achieve a cell density of 10,000 cells/cm² was calculated and split in equal volumes in two separate centrifuge tubes. Cell pellets were then obtained by centrifuging the cell suspensions at 200g for 5 minutes. Each cell pellet was then resuspended in a working solution of Cell Tracker™ Red CM-Dil (working concentration of 20 µM) and Cell Tracker™ Green CMFDA (working concentration of 20 µM), respectively, and incubated in the dark at 37°C for 40 minutes. After incubation, the Cell Tracker™ working solutions were removed by centrifugation and replaced with fresh pre-warmed growth medium. The fluorescently-labelled cells were then seeded at 10,000 cells/cm² on the pre-conditioned microcarriers in spinner flasks. Combinations of two different microcarriers (Plastic and Cytodex-1) and two different fluorescent dyes were used for this study. The labelled cells were allowed to attach and proliferate for 48 h in agitated conditions, in the dark, at 37°C and 5% CO₂. After 48 h in culture, the different microcarrier-labelled cell suspensions were mixed in a 50:50 proportion and the culture was continued for an additional 2 days. The mixed microcarrier-labelled cell suspensions were then imaged on the epi-fluorescence microscope mentioned above.

2.6 Statistical analysis

For comparison between the two data sets comparing the mean specific growth rates, statistical significance was determined by using the Student's two-tailed t-test. Significance was determined at $p < 0.05$ (*) or $p < 0.01$ (**). GraphPad Prism 6 (California, USA) was used for the statistical analysis

3. Results and Discussion

The aim of this work was to demonstrate, both quantitatively and qualitatively, the phenomenon of bead-to-bead transfer of hMSCs on microcarriers under different culture conditions including cells from three BM-hMSC donors and under different culture medium conditions, FBS, HPL and SFM. An additional aim was to investigate the potential for process intensification through cells-on-beads inoculation, where 10% of near-confluent microcarriers were used to inoculate a fresh microcarrier culture.

Cell-microcarrier contact in bead-to-bead hMSC transfer is very likely to be influenced by a combination of factors such as donor cell line, medium composition, attachment substrate (*i.e.* microcarriers) and the process itself. The process employed with respect to the feeding regime, surface area provided or agitation intensity was kept constant during this study using a protocol that had been thoroughly tested and optimised for the successful expansion of bone-marrow derived hMSCs in spinner flasks in different medium conditions (Heathman *et al.* 2015a; Rafiq *et al.* 2016). To account for donor variability, hMSCs from three donors were employed for this study.

3.1 *Qualitative demonstration of bead-to-bead transfer*

To qualitatively demonstrate hMSC migration from one microcarrier to another, two different methods were employed. The first method made use of commercially available microcarriers with different visual properties, while the second method consisted of fluorescently-labelling cells, followed by tracking their movements in culture.

Prior work with multiple types of microcarriers in a microcarrier screening study (Rafiq *et al.* 2016) showed that BM-hMSCs would attach and grow on most of the microcarriers albeit with varying degrees of success on each. It was also found that certain microcarriers were visually distinctive compared to others, for example the Cytodex-1 microcarriers are translucent in comparison to the

solid Plastic P102-L microcarriers, and the Hillex microcarriers absorb the phenol red present in the medium and therefore turn a dark red colour. To demonstrate the phenomenon of hMSC bead-to-bead transfer qualitatively, the BM-hMSCs were initially cultured exclusively on Plastic P102-L microcarriers for a period of 6 days, after which fresh Hillex microcarriers (Figure 1A-C) or fresh Cytodex-1 microcarriers (Figure 1D-F) were aseptically added to the culture without stopping agitation of the culture at any point.

As is indicated by the arrows in the figures, it is evident that BM-MSCs were attached to both the Plastic P102-L microcarriers (solid white) and the Hillex (dark red) microcarriers (Figure 1A-C). Similarly, cells were attached to the Plastic P102-L and Cytodex 1 (translucent white) microcarriers (Figure 1D-F). The BM-hMSCs were found to be growing on microcarriers of both types as either cell-microcarriers aggregates, as well as on individual Plastic P102-L, Hillex and Cytodex-1 microcarriers (Figure 1A-F). This observation is consistent with both the bridging and cell migration methods of bead-to-bead transfer noted by Leber and colleagues in their studies using an immortalised hMSC line (Leber et al. 2017). It is worth noting that following the addition of the second type of microcarrier to the culture vessel, at no point was the agitation stopped and N_{JS} was maintained throughout. These findings therefore provide a qualitative demonstration of bead-to-bead transfer during a continuing culture, whereby hMSCs have attached and proliferated on 'fresh' microcarriers days after the addition to the culture.

To demonstrate that these were indeed viable cells attached to both types of microcarriers (Plastic P102-L and Hillex), samples were stained with calcein AM (viable stain, green) and ethidium homodimer (non-viable stain, red) and assessed under a fluorescent microscope (Figure 2). It is evident that the cells which are attached to the microcarriers in phase contrast images are indeed viable cells given the extent of viability staining exhibited in Figures 2B and 2D. Similar findings

were found with the Plastic P102-L and Cytodex-1 pairing and the results depicted in Figure 2 are representative of that combination of microcarriers too.

To further demonstrate the bead-to-bead transfer qualitatively, the cells were fluorescently labelled with two different fluorescent dyes prior to inoculation on the microcarriers. These Cell Tracker™ dyes are virtually non-fluorescent until cytosolic esterases found only inside living cells cleave off the acetate moiety, releasing a brightly fluorescent product. Moreover, these reagents pass freely through cell membranes, but once inside the cell, are transformed into cell-impermeant reaction products. Further, the fluorescent probes are inherited by the daughter cells and are not transferrable to adjacent cells in a population. As such, the Cell Tracker™ Red-labelled cells were cultured initially only onto Plastic microcarriers (solid) and the Cell Tracker™ Green-labelled cells cultured initially only onto the Cytodex-1 microcarriers (transparent) for 2 days. After combining the two different types of microcarriers, there was evidence that Cell Tracker™ Red-labelled cells have migrated from Plastic onto Cytodex-1 microcarriers (white arrows) and *vice versa* (yellow arrows) (Figure 3A-3D). Notably, the migrated cells exhibited a more rounded morphology than these adherent cells normally do. During mitosis, adherent cells undergo a cycle of detachment and re-attachment (Yamakita et al. 1999) so this could be indicating active cell growth on the 'new' microcarrier that the cell has migrated to or it could be an indication that following detachment from another microcarrier during mitosis, the cell had not yet fully adhered at the time of imaging.

3.2. Quantitative demonstration of bead-to-bead transfer

To quantitatively demonstrate bead-to-bead transfer, BM-hMSC1 and BM-hMSC2 were grown for 6 days on Plastic P102-L microcarriers in either FBS-based or SFM culture conditions (Figure 4, Culture 1), after which, 10% of the microcarriers (with cells attached) were seeded into a fresh

culture vessel with 90% fresh Plastic P102-L microcarriers (Figure 4, Culture 2). Upon inoculation of the microcarriers with cells attached, the vessel was immediately agitated at N_{JS} . This was repeated for a third time (Figure 4, Culture 3).

With each consecutive culture, it is noteworthy that the initial lag phase is reduced and with each culture, the final cell density obtained by the end is greater than the previous. This is evident from both the cell density data across the three cultures (Figure 4) and the population doublings and doubling time data (Table 1). For FBS cultures, the final cell density was found to be $\sim 1.6 \times 10^5 \pm 0.15$ cells/mL obtained with Culture 1, $2.01 \times 10^5 \pm 0.06$ cells/mL in Culture 2 and $\sim 2.15 \times 10^5 \pm 0.08$ cells/mL obtained in Culture 3 for hBM-MSC1 and a similar trend was found for hBM-MSC2 (Figure 4). For SFM cultures, the cell yield was higher in all cases, but the same trend of reduced lag phase and increasing yield within the same time period (6 days) was noted. Moreover, in all cases, viability was $> 95\%$, indicating favourable culture conditions in the vessel for the growth of hMSCs throughout the course of each culture. These findings indicate that a 10% concentration of near-to-confluent microcarriers was not only sufficient to inoculate a new vessel containing 90% fresh microcarriers, but decreased the lag phase of the new culture and increased the overall yield.

The findings would appear to demonstrate bead-to-bead transfer given the fact that a 10% microcarrier concentration (with cells attached) was able to successfully inoculate a culture vessel with fresh microcarriers, with the culture eventually reaching a point of near-confluency prior to termination of the culture. Similar findings were observed by Ohlson *et al.* (1994) and Wang and Ouyang (1999) for adherent CHO cells who performed similar inoculation studies with near-confluent and fresh microcarriers. Moreover, Ohlson *et al.* (1994) also observed an increase in the overall cell yield and a reduced lag phase, similar to what was found in this study. In addition to investigating a 10% addition of near-confluent microcarriers to a culture containing fresh microcarriers, Ohlson *et al.* (1994) also investigated inoculating a 25% concentration of near-

confluent microcarriers to a fresh culture and found that this resulted in a further reduced lag phase and an even greater cell density than the 10% near-confluent microcarrier addition. Although a 25% concentration of near-confluent microcarriers were not investigated in our study, that earlier finding indicates that an important process development activity might be to establish the optimal concentration of near-confluent microcarriers to inoculate into fresh bioreactor systems. This analysis will undoubtedly be a trade-off between the number of bioreactors to be seeded and the desired culture growth kinetics (similar to splitting a near-confluent T-flasks into multiple T-flasks). In a separate study, Badenes et al. (2017) demonstrated bead-to-bead transfer with hPSCs, where it was found that this technique facilitated an increase in viable cell number and a concomitant significant increase in fold expansion within a 15 day culture.

To further quantitatively demonstrate the phenomenon of bead-to-bead transfer, a fluorescence microscopy-based manual cell counting technique was adopted to highlight the difference between two inoculation methods. One (the control) used the well-established technique of cells in free suspension and the other, used inoculation of an equivalent number of cells attached to 10% of the beads. With these two methods, one would expect there to be a distinct difference in how the distribution of the hMSC population across the microcarriers changed over the culture period. For the control method using cells in suspension, the expectation would be very few cells per microcarrier at the start of the culture and with time as the hMSCs proliferated, this cell distribution would shift to higher numbers of cells per microcarriers. With the inoculation via the bead-to-bead transfer method, the expectation would be that the cell distribution splits into two distinct populations at the start of the culture, one population with a lot of cells, being the microcarriers added with cells attached, and the other being empty microcarriers. As the hMSCs proliferate and bead-to-bead transfer occurs, one would expect these two populations to merge over the course of the growth phase, finally producing similar distributions at the end of the cultures.

The data presented (Figure 5A) reflects the expected trends, particularly in the case of inoculating cells in suspension as is traditionally done (the control) where the number of cells per bead began with many beads without cells at day zero with 75% of beads with 1 to 5 cells on day 1. The maximum number of cells per bead steadily increased until eventually at day 5, the maximum number of beads were those approaching or at confluence with between 5 to 15 cells per bead on the remainder.

With the inoculation via cells on beads followed by bead-to-bead transfer (Figure 5B), few beads have many cells (some approaching confluence) and the remainder have none on day 0. With the passage of time up to day 5, fewer and fewer beads are without cells and the number of cells is approximately the same on the other beads and steadily increases. At the end (day 5), a very high proportion of the beads are approaching or have reached confluence.

3.3 Comparison of inoculation techniques

Having established that BM-hMSCs can transfer to freshly added microcarriers and that a 10% concentration of near-confluent microcarriers could act as a sufficient inoculum for a fresh culture, to further prove the efficacy of the new approach, it was decided to include cells from a third donor (BM-hMSC 3) to supplement the two used in the experiments establishing bead-to-bead transfer (BM-hMSC 1, BM-hMSC 2). Also a third culture medium (HPL-based) was included. The results inoculating with these three donor cells on Plastic P102-L microcarriers and three different medium are summarised in Figure 6 (FBS-based), Figure 7 (SFM) and Figure 8 (HPL-based). The results are also compared to our previous approach of free suspension inoculation as control (Rafiq et al. (2016); Heathman et al. (2015a)). Though the results differed between the different medium, the data were very similar for each donor cell. Thus, the data presented in Figures 6-9 illustrate a typical result for any of the cells in that specific medium conditions.

There was a significant improvement in performance between the two approaches in all culture medium conditions (FBS, HPL and SFM) throughout the course of the culture, resulting in a higher cell density at the end of the culture when utilising cells-on-beads inoculation. For the FBS-based culture, the cell density was $\sim 2.0 \times 10^5$ cells/mL compared to $\sim 1.5 \times 10^5$ cells/mL respectively (Figure 6A) and for SFM, the final cell densities were $\sim 4.6 \times 10^5$ cells/mL, compared to $\sim 3.4 \times 10^5$ cells/mL respectively (Figure 7A). A similar trend was found for HPL, where the values were $\sim 3.6 \times 10^5$ and $\sim 3.0 \times 10^5$ cells/mL respectively (Figure 8A). This increase in cell yield is associated with a reduced lag phase for all cultures using the cells-on-beads inoculation method, enabling the cells to reach the higher growth phase of the culture quicker in comparison to the control. This more rapid achievement of fast growth is illustrated in Figures 6-8B and Figures 6-8C. Figures 6-8 B, shows that for FBS, SFM and HPL cultures, the cells' mean specific growth rate was significantly higher between days 0-2 with the cells-on-beads inoculation when compared to the controls (Figures 6-8C). After that, the growth rate for each method reached parity between days 2-4. These figures also make clear that there was a reduced lag phase with cells-on-beads inoculation.

The specific glucose consumption rate along with the specific lactate and ammonia production rates were also calculated for each culture (Figure 9). For FBS-based culture, the specific glucose consumption rate during pre-culture for both the inoculation via cells-on-beads and the control method was ~ 19 pmol.cell⁻¹.d⁻¹. Then between days 0-2 post inoculation, these values sharply rose to ~ 38 pmol.cell⁻¹.d⁻¹ for the inoculation via cells-on-beads and was even higher (~ 45 pmol.cell⁻¹.d⁻¹) for the control. Between days 2-4, the glucose consumption rates have settled back down to similar values of ~ 20 pmol.cell⁻¹.d⁻¹ for both methods. This increase in specific glucose consumption rate during the first 2 days of culture was mirrored by an increase in the specific lactate and ammonia production rates during the same time phase. A distinct trend could be seen across each

medium type, where in general an increase in specific consumption/production rates over the first 2 days of culture was significantly reduced when using the inoculation via cells-on-beads instead of the control. It must be noted that all rates were significantly reduced when using a serum-free medium.

This trend suggested that using inoculation by cells-on-beads resulted in a reduced impact on the metabolic activity of the hMSCs during the early phase of culture that can likely be attributed to the production of extracellular components required for attachment when using the control method. Although it has not been explored here, this highlights a potential impact of multiple trypsinisation steps on the potency or long-term proliferative capability of hMSCs which could be avoided by adopting the inoculation via cells on beads followed by bead to bead transfer.

With respect to characterisation, the BM-hMSCs, both pre-inoculation and post-harvest were characterised with respect to cell identity (immunophenotypic expression) and functionality (differentiation potential). In all cases (different culture medium conditions and inoculation methods), the BM-hMSCs were found to be in accordance with the ISCT criteria (Dominici *et al.* 2006). Representative data is shown for the condition wherein the BM-hMSCs were cultured in SFM and inoculated using cells-on-beads. However similar findings were found for all other conditions (data not shown). The data demonstrates that there was > 95% co-expression of CD105, CD90 and CD73 and less than 2% positive for the negative markers CD34 and HLA-DR for the harvested BM-hMSCs (Supplementary Figure 1). Moreover, the cells were analysed for the retention of their tri-lineage differentiation potential towards the osteogenic, chondrogenic and adipogenic pathways. Again, in all cases, the BM-hMSCs were found to retain their differentiation potential (Supplementary Figure 1).

The impact of this study is particularly relevant to the production phase for hMSC manufacture, where the need for large numbers of cells within a limited time period may be necessary. This

approach allows for multiple bioreactors to be inoculated and operated in parallel from a small percentage of near-confluent microcarriers, which lends itself well to an allogeneic cell therapy. This production approach is akin to T-flask based culture, where a near-confluent T-flask is 'split' into other T-flasks to facilitate the progression of culture. The new approach offers a significant advantage in that the scale of a stirred-tank bioreactor is significantly larger than that of a T-flask and therefore so is the yield of cells that can be obtained. Combined with the ability to increase surface area during the course of the culture through the addition of fresh microcarriers, this approach facilitates significant expansion potential. Unlike a traditional seed train process from primary cells, this process could enable cells to be cultured at relatively low population doublings in multiple reactors before either being frozen down as a part of a master/working cell bank, or indeed, for the final manufacture of cells.

4. Conclusions

Bead-to-bead transfer is widely believed to occur in anchorage-dependent microcarrier cultures. Unlike previous studies investigating this phenomenon, this study describes the approach taken to demonstrate bead-to-bead transfer, both qualitatively and quantitatively, for three different BM-hMSC donors and under different culture medium conditions, FBS, HPL and SFM. Qualitative demonstration of the phenomenon was achieved by using microcarriers with distinctive visual properties and cell-tracking fluorescent reagents where it was demonstrated that freshly added microcarriers to an existing culture were soon populated by BM-hMSCs, with examples of cells bridging between different microcarriers, and in other instances cells attaching and proliferating on the freshly added microcarriers in isolation. Quantitative demonstration of the phenomenon was established by the process of taking 10% of near-confluent microcarriers to inoculate a fresh microcarrier culture (cells-on-beads inoculation), where it was found that bead-to-bead transfer not only occurred, but that this inoculation method, in contrast to the control (where cells were inoculated *via* free suspension), increased cell growth rate and overall cell yield. The increase in cell yield was associated with a reduced lag phase for all cultures using the cells-on-beads inoculation method, enabling the cells to reach the higher growth phase of the culture quicker in comparison to the control. It was also demonstrated that the cell identity and differentiation potential was retained in all culture medium conditions and with both inoculation methods. Cells-on-beads inoculation is clearly an effective means of process intensification and can have a significant impact on reducing overall expansion times and therefore the costs of BM-hMSC and other anchorage-dependent cell therapies.

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Conflict of interest

The authors declare no financial or commercial conflict of interest.

Tables:

Table 1: Population doublings and doubling time data for BM-hMSC1 and BM-hMSC2 in FBS and SFM across 3 consecutive cultures

BM-hMSC line	Medium condition	Culture number	Doubling time (h)	Population Doublings
BM-hMSC1	FBS	1	56.4	2.55
		2	51.0	2.82
		3	48.1	2.99
	SFM	1	40.7	3.54
		2	38.2	3.77
		3	37.0	3.89
BM-hMSC2	FBS	1	58.6	2.46
		2	52.8	2.73
		3	51.4	2.80
	SFM	1	42.6	3.38
		2	41.6	3.46
		3	40.3	3.57

Figure captions

Figure 1: Phase contrast images of the bead-to-bead transfer between the different types of microcarriers, Plastic P102-L (white with black rim), Hillex (red) and Cytodex-1 (translucent white). The white arrows indicate cell-microcarrier aggregates between two different microcarriers. The black arrows indicate single cells attached to a specific microcarrier. Representative images from BM-hMSC1 (A, B, E, F), BM-hMSC2 (C, D, G, H) are illustrated. Figures were taken at day 6 after inoculation (3 days after adding the second type of microcarrier).

Figure 2: Phase contrast (A; C) and Live/Dead staining (B; D) images of the same microcarriers and cells demonstrating the bead-to-bead transfer. Representative images from BM-hMSC1 (A,B) and BM-hMSC2 (C,D) are depicted with similar findings across both donors used. Figures were taken at day 6 after inoculation (3 days after adding the second type of microcarrier). The microcarriers used were Plastic (white with black rim) and Hillex (red).

Figure 3: Fluorescent and bright field images showing Cell Tracker Red-labelled cells migrated from Plastic (solid) to Cytodex-1 (transparent) microcarriers (white arrows) and Cell Tracker Green-labelled cells migrated from Cytodex-1 to Plastic microcarriers (yellow arrows). (A) Bright field image; (B) Cell Tracker-Green; (C) Cell Tracker-Red; (D) Merged. Scale bar represents 100 μm .

Figure 4: Quantitative demonstration of bead-to-bead culture in FBS and SFM cultures with BM-hMSC1 and BM-hMSC2 donor cells cultured on Plastic P102-L microcarriers. Data show mean \pm SD, n = 3.

Figure 5: Cell distribution on microcarriers when using (A) the standard cell inoculation method or (B) the cells-on-beads inoculation method. The gradation in colour (from light to dark) represents the numbers of cells on microcarriers. Data represents BM-hMSC3 cells cultured on Plastic P102-

L microcarriers in HPL. The data is representative of findings from the other donor cells and cultured in the other medium compositions.

Figure 6: Comparison of BM-hMSC1 growth on Plastic P102-L microcarriers in a FBS-containing medium when using the standard cell inoculation (control) or cells-on-beads inoculation, where the data represents, (A) the cell density (cells/mL) over 7 days, (B) the mean specific growth rate (d^{-1}) over the course of the culture and (C) a logarithmic plot of the cell density across the culture period. Data show mean \pm SD, $n = 3$. Significance was determined at $p < 0.05$ (*) or at $p < 0.01$ (**).

Figure 7: Comparison of BM-hMSC2 growth on Plastic P102-L microcarriers in the SFM when using the standard cell inoculation (control) or cells-on-beads inoculation, where the data represents, (A) the cell density (cells/mL) over 7 days, (B) the mean specific growth rate (d^{-1}) over the course of the culture and (C) a logarithmic plot of the cell density across the culture period. Data show mean \pm SD, $n = 3$. Significance was determined at $p < 0.05$ (*) or at $p < 0.01$ (**).

Figure 8: Comparison of BM-hMSC3 growth on Plastic P102-L microcarriers in a HPL-containing medium when using the standard cell inoculation (control) or cells-on-beads inoculation, where the data represents, (A) the cell density (cells/mL) over 7 days, (B) the mean specific growth rate (d^{-1}) over the course of the culture and (C) a logarithmic plot of the cell density across the culture period. Data show mean \pm SD, $n = 3$. Significance was determined at $p < 0.05$ (*) or at $p < 0.01$ (**).

Figure 9: Metabolite data for the cultures comparing the standard cell inoculation (control) and cells-on-beads inoculation across the different culture medium conditions, FBS, HPL and SFM. (A-C) represents the mean specific glucose consumption rate ($\text{pmol}\cdot\text{cell}^{-1}\cdot\text{d}^{-1}$), the mean specific lactate production rate ($\text{pmol}\cdot\text{cell}^{-1}\cdot\text{d}^{-1}$) and the mean specific ammonia production rate ($\text{pmol}\cdot\text{cell}^{-1}\cdot\text{d}^{-1}$) for the FBS culture. (D-F) represents the mean specific glucose consumption rate ($\text{pmol}\cdot\text{cell}^{-1}\cdot\text{d}^{-1}$), the mean specific lactate production rate ($\text{pmol}\cdot\text{cell}^{-1}\cdot\text{d}^{-1}$) and the mean specific ammonia production

rate ($\text{pmol}\cdot\text{cell}^{-1}\cdot\text{mL}^{-1}$) for the HPL culture. (G-I) represents the mean specific glucose consumption rate ($\text{pmol}\cdot\text{cell}^{-1}\cdot\text{mL}^{-1}$), the mean specific lactate production rate ($\text{pmol}\cdot\text{cell}^{-1}\cdot\text{d}^{-1}$) and the mean specific ammonia production rate ($\text{pmol}\cdot\text{cell}^{-1}\cdot\text{d}^{-1}$) for the SFM culture. Data show mean \pm SD, n = 3.

Supplementary Figure 1: Immunophenotypic expression for BM-hMSC1 (A) and differentiation of the BM-hMSC1 toward the (B) adipogenic, (C) osteogenic and (D) chondrogenic lineages. All data are of cells cultured on Plastic P102-L microcarriers. Representative data was obtained for all other cell donors and in all other medium conditions.

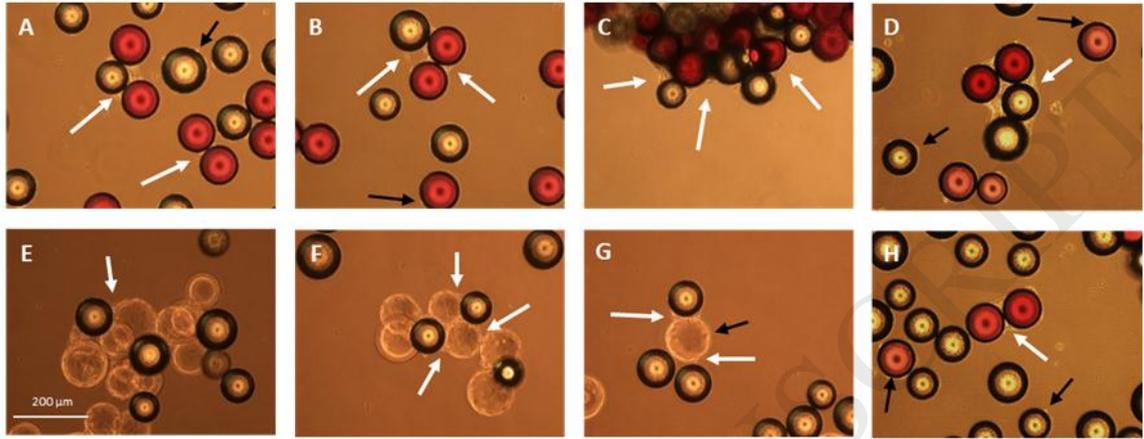
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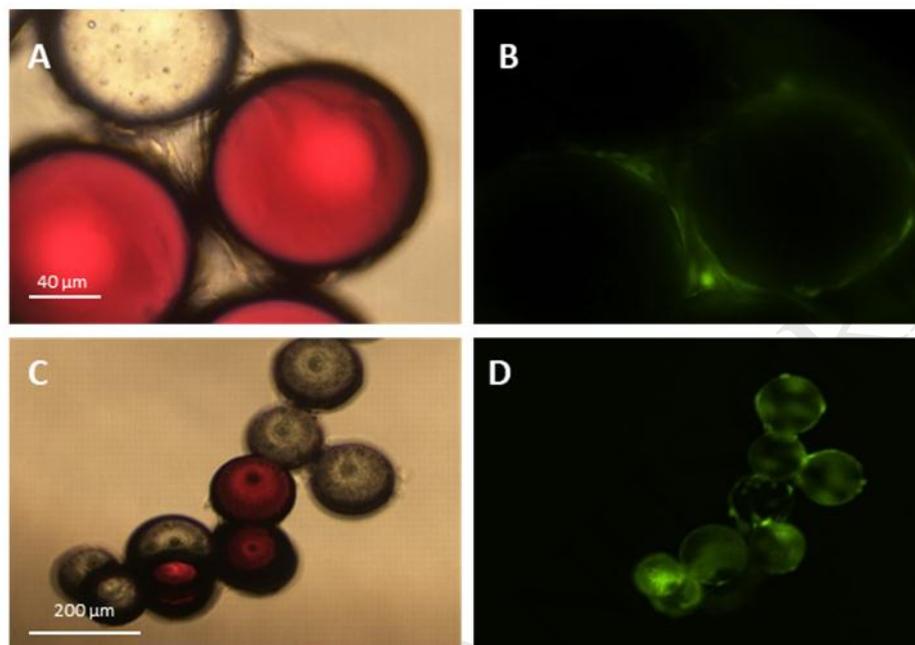
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Figure Caption

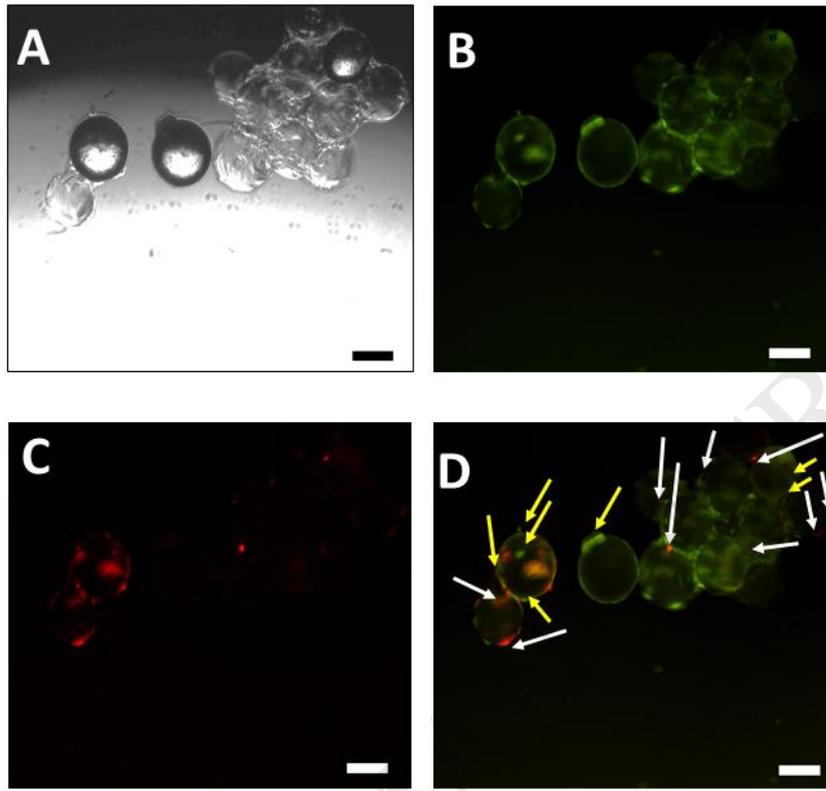
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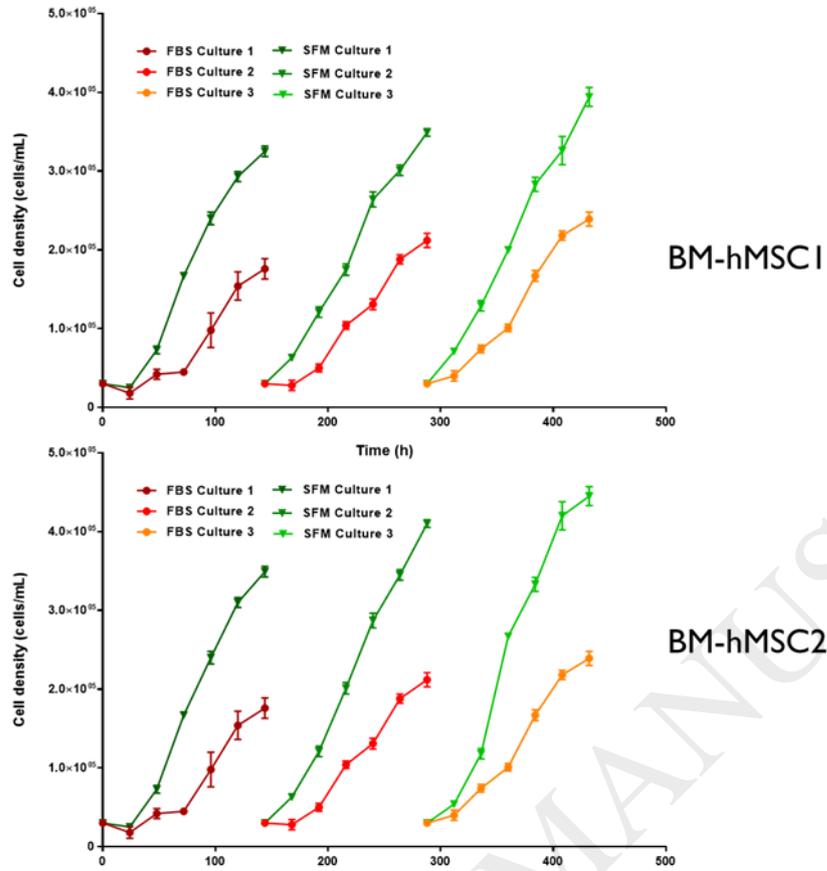
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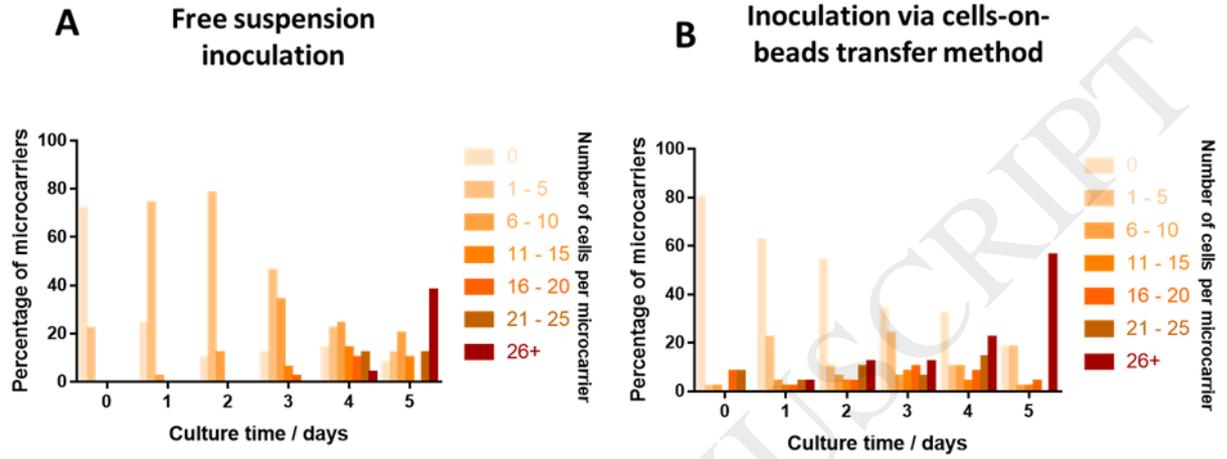
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Figr-5



Figr-6

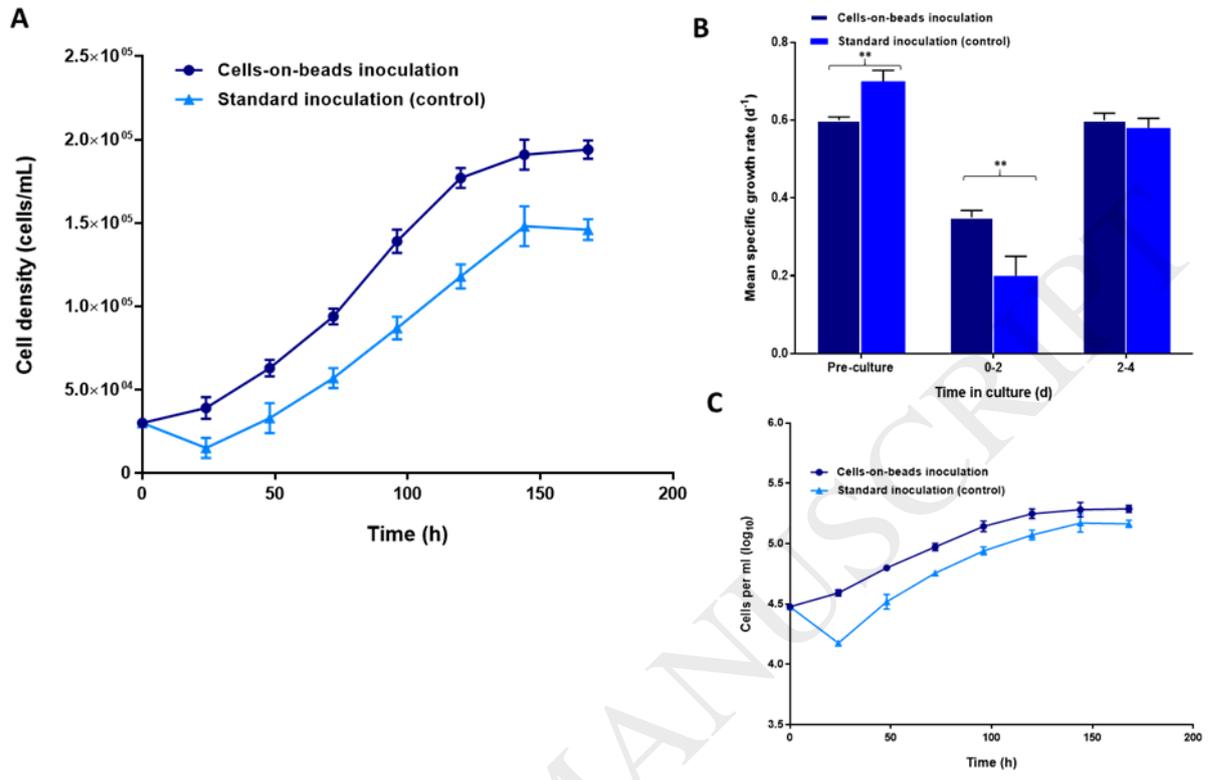


Fig-7

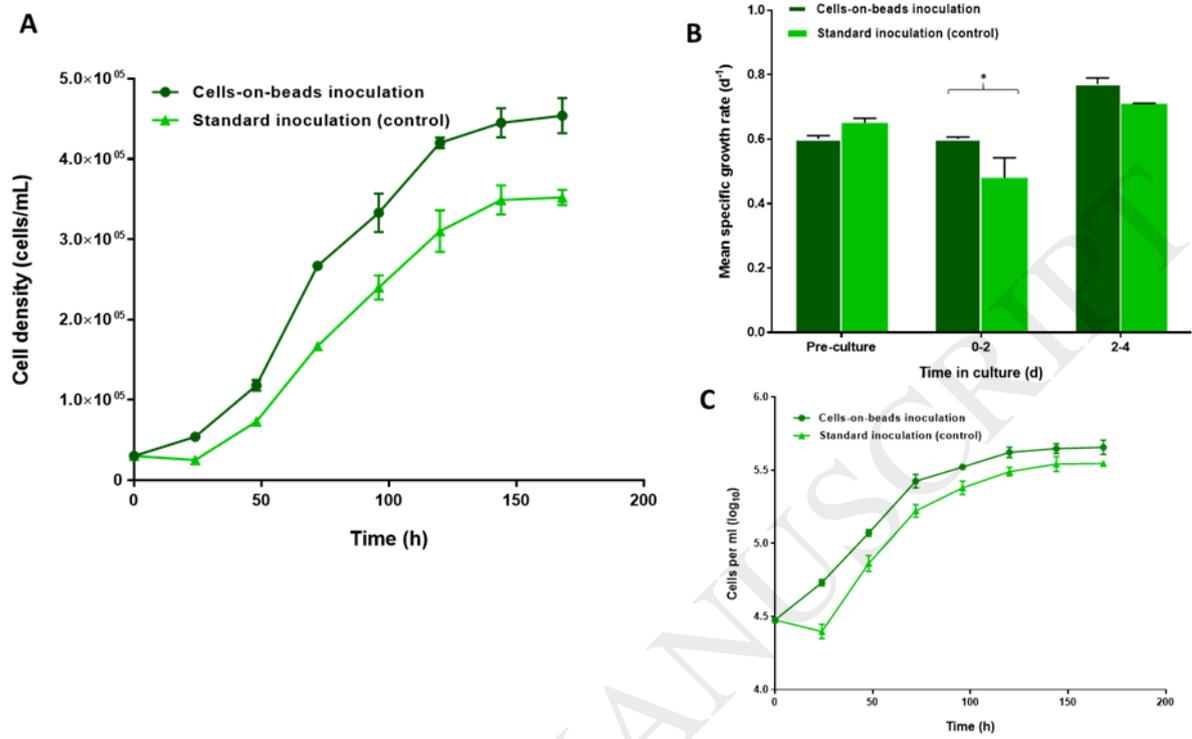


Fig-8

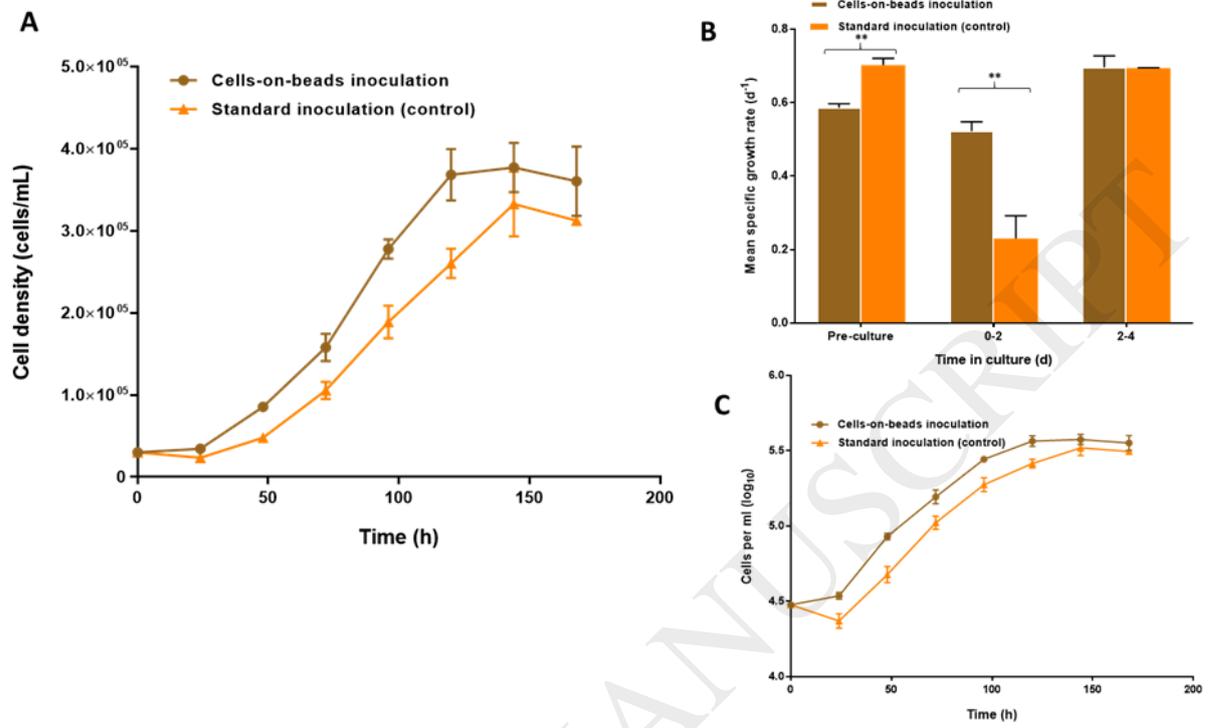


Fig-9

