# A Duration-Dependent Interaction Between High-Intensity Light and Unrestricted Vision in the Drive for Myopia Control

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Citation: Biswas S, Muralidharan AR, Betzler BK, et al. A duration-dependent interaction between high-intensity light and unrestricted vision in the drive for myopia control. *Invest Ophthalmol Vis Sci.* 2023;64(3):31. https://doi.org/10.1167/iovs.64.3.31 **PURPOSE.** To evaluate the duration-dependent and synergetic impact of high-intensity light (HL) and unrestricted vision (UnV) on lens-induced myopia (LIM) development in chickens.

**M**ETHODS. Myopia was induced in one eye in chicks (10 groups, n = 126) from day 1 posthatching (D1) until day 8 (D8) using -10 diopter (D) lenses. Fellow eyes remained uncovered as controls. Nine groups were exposed daily to 2, 4, or 6 hours of HL (15,000 lux), UnV (removal of -10 D lens), or both (HL + UnV). One group served as the LIM group without any interventions. Ocular axial length (AL), refractive error, and choroidal thickness were measured on D1, D4, and D8. Outcome measures are expressed as interocular difference (IOD = experimental eye – control eye) ± SEM.

**R**ESULTS. By D8, LIM increased AL  $(0.36 \pm 0.04 \text{ mm})$ , myopic refraction  $(-9.02 \pm 0.37 \text{ D})$ , and choroidal thinning  $(-90.27 \pm 16.44 \mu\text{m})$  in the LIM group (all, P < 0.001). Compared to the LIM group, exposure to 2, 4, or 6 hours of HL, UnV, or HL + UnV reduced myopic refraction in a duration-dependent manner, with UnV being more effective than HL (P < 0.05). Only 6 hours of HL + UnV (not 2 or 4 hours) prevented LIM and was more effective than UnV (P = 0.004) or HL (P < 0.001) in reducing myopic refraction and more effective than HL (P < 0.001) in reducing axial elongation.

**C**ONCLUSIONS. Daily exposure to 2, 4, or 6 hours of HL, UnV, or HL + UnV reduced lensinduced myopic refraction in a duration-dependent manner in chickens. Only 6 hours of HL + UnV completely stopped LIM development. The synergetic effect of HL and UnV is dependent on the duration of the interventions.

Keywords: myopia, animal model, unrestricted vision, optical defocus, high-intensity light, axial length, choroid, outdoor time

**M** yopia, also known as near-sightedness, is one of the most prevalent eye conditions worldwide and has become a public health concern, particularly in East and Southeast Asia.<sup>1</sup> Axial myopia, the most common form of myopia, involves excessive globe elongation during ocular development, resulting in a mismatch between the axial length (AL) and optical power of the eye.<sup>2</sup> While the refractive error associated with myopia can be corrected, high myopia is associated with sight-threatening pathologies,

including retinal detachment, glaucoma, and myopic maculopathy.<sup>3</sup>

Environmental factors alongside genetics are known to play a significant role in the development of myopia.<sup>4</sup> Increased outdoor activity in childhood reduces the risk of developing myopia,<sup>5–7</sup> independent of physical activity.<sup>8</sup> This protective impact of outdoor time may be due to a combination of factors, including the intensity and spectral composition of sunlight,<sup>9,10</sup> the increased spatial frequency

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and higher image contrast encountered outdoors,<sup>11,12</sup> and the significant differences in the pattern of retinal defocus generated outdoors compared with indoors,<sup>11,13</sup> associations that are still under investigation in animal models of myopia.<sup>14</sup> Because time spent outdoors involves all of these factors simultaneously, the relative protective contribution of each and interaction between them are unknown. Animal models offer an opportunity to explore such associations.

The most commonly used myopia induction methods in animal models are form-deprivation myopia (FDM), in which translucent diffusers are fitted to obscure the animal's vision, and lens-induced myopia (LIM), in which hyperopic defocusing lenses (i.e., negative powered lenses) are fitted to place the focal plane behind the retina and induce axial ocular growth.<sup>15</sup> While it has been suggested that FDM and LIM have different underlying mechanisms of action,<sup>16-18</sup> more evidence is needed to support these claims given the biochemical,<sup>19-21</sup> genetic,<sup>22</sup> and anatomic<sup>23</sup> similarities between these models of myopia induction, in addition to similarities in their response to pharmacologic agents.<sup>24</sup>

Exposure to high-intensity light (HL; 10,000-25,000 lux per day) can be effective in alleviating FDM in various animal models.<sup>25-28</sup> In addition, Ashby et al.<sup>29</sup> reported that in chicks, the suppression of FDM is even more pronounced when HL was combined with normal vision (diffuser removal) for 15 min/d, compared to experimental groups exposed to either HL (15,000 lux) or diffuser removal alone. The authors postulated that dopamine (DA) release in the chick retina had a graded response to increasing illumination due to diffuser removal, resulting in greater inhibition of axial ocular growth at higher illumination levels.<sup>29</sup> In studies utilizing LIM, exposure to HL without the defocusing lens removal or unrestricted vision (UnV) significantly slowed LIM development in chicks (15,000 lux for 5 h/d)<sup>30</sup> but not in monkeys (25,000 lux for 6 h/d),<sup>31</sup> while exposure to brief periods (1-3 hours) of UnV per day reduced the extent of LIM development in tree shrews,<sup>32,33</sup> marmosets,<sup>34</sup> and chicks.<sup>35</sup> While LIM and FDM may yield different ocular mechanisms,<sup>16,17</sup> lens removal for LIM and normal vision in case of FDM may operate through different pathways. In LIM, UnV may act through a change in optical and visualspatial information, whereas normal vision in FDM additionally involves neuromodulations associated with increased retinal illumination.36-38

To our knowledge, the dose-dependent and synergetic effects of HL, UnV, and HL + UnV are yet to be investigated in LIM models. This study investigates the interactive impact of different combinations and durations of HL and UnV on ocular axial elongation, refractive error development, choroidal thickness, and other ocular parameters in a chicken model of LIM.

# **METHODS**

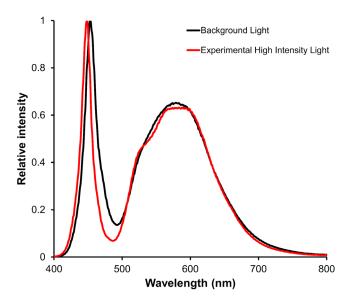
#### **Animals and Enclosure**

All animals used in this study were treated in accordance with the ARVO Statement for the Use of Animals in Ophthalmic and Vision Research. The study protocol was approved by the Institutional Animal Care and Use Committee (IACUC 2019/SHS/1479) of the Singapore Experimental Medicine Centre (SEMC). The SEMC is accredited by the Association for Assessment and Accreditation of Laboratory Animal Care International.

In total, 126 one-day-old chicks (mixed Golden Comet/White Leghorn strain) were provided by the National Large Animal Research facility and randomly assigned into 10 groups of 11 to 13 animals each. Animals were raised for 9 days in a 75-cm (length) × 55-cm (width) × 43-cm (height) custom-built enclosure designed to hold two high-intensity light-emitting diode (LED) light fixtures. The enclosure walls were fitted with a square wave grating of a repeated sequence of light and dark bars as accommodative cues. The spatial frequency of the stripes ranged between 0.01 and 0.42 cycles/degree depending on the location of the animal within the enclosure. The same visual environment was maintained across experimental groups to avoid differences in accommodative responses that may affect emmetropization in chicks.<sup>39</sup> The chicks were fed ad libitum and were raised under a 12/12-hour light-dark cycle from 7 AM to 7 PM. The temperature within the enclosure was maintained at 28°C to 32°C. Light and temperature patterns were monitored using a HOBO Pendant data logger (UA-022-64; ONSET, Bourne, MA, USA). At the end of experiment on day 9, chicks were sedated with a mixture of 0.2 mL/kg ketamine and 0.1 mL/kg xylazine and euthanized with an overdose of sodium pentobarbitone to the heart.

#### **Background and Experimental Light Setup**

All chicks were reared under background lighting (150 lux) during the 12/12-hour light-dark cycle. This was achieved using six strips of ultra-bright LEDs (4000K, 2NFLS-NW LED; Super Bright LED, Inc, St. Louis, MO, USA) fixed over the enclosure. For HL, four LED panels of 64 LEDs each (4000K; USHIO Lighting, Singapore) were used to deliver an average of 15,000 lux when measured in different angles of gazes in the enclosure (up, down, left, right, front, back) (Fig. 1). The light fixtures were controlled using a programmable Helvar DIGIDIM 910 router (Helvar, Dartford Kent, UK). Light levels and spectra were measured using a calibrated radiometer and spectroradiometer (ILT5000 and ILT950; International Light Technologies, Peabody, MA, USA).



**FIGURE 1.** Relative spectral power distribution of the background light (LED, 4000K, 2NFLS-NW LED; Super Bright LED, Inc.) and experimental high-intensity light (LED, 4000K; USHIO Lighting).

TABLE. Details on Experimental Groups and Interventions

Experimental Group	Duration of Intervention, h	N	Experimental Eye	Control Eye	<b>Experimental Interventions</b>	
					High-Intensity Light Status (15,000 Lux)	Lens Status
LIM	0	13	-10 D	No lens	Off	Not removed
HL	2	13	-10 D	No lens	On	Not removed
	4	13	-10 D	No lens	On	Not removed
	6	13	-10 D	No lens	On	Not removed
UnV	2	12	-10 D	No lens	Off	Removed
	4	13	-10 D	No lens	Off	Removed
	6	13	-10 D	No lens	Off	Removed
HL + UnV	2	13	-10 D	No lens	On	Removed
	4	12	-10 D	No lens	On	Removed
	6	11	-10 D	No lens	On	Removed

# **Myopia Induction**

Monocular myopia was induced in all chicks from day 1 posthatching (D1) until day 8 (D8) using custom-built concave defocusing lenses (power:  $-10 \pm 0.5$  diopters [D], total diameter: 12.5 mm, optic zone diameter: 10 mm, base curve: 6.68 mm; La SER Eye Jewelry, Port St. Lucie, FL, USA). The lens was fitted randomly to one eye of the chick using a custom-built three-dimensional printed lens holder. Considering the diameter of the optic zone (10 mm), an estimated vertex distance (defocusing lens to cornea apex) of 3 mm, and a calculated posterior nodal point to defocusing lens distance of 4.49 mm on D1 in chicks, the open viewing visual angle was estimated to be  $\sim$ 76.5 degrees. However, these calculations did not take into account changes in pupil size and may have underestimated the open viewing visual angle.<sup>40</sup> The lens holder can be clipped on or off a separate base piece that was glued to the down surrounding the chick's eye. These lens holders ensured the secure positioning of the lenses on the eyes of the animals and allowed for easy and fast removal during cleaning and light exposure. Lenses were worn for 8 days and were cleaned three times/d to ensure their optical clarity. The fellow eye was kept uncovered and used as a within-animal control.

# **Experimental Groups**

All 10 groups were subjected to monocular LIM. Nine groups were subjected to different interventions, including either transient HL (15,000 Lux), UnV, or a combination of HL and UnV for one of four durations/d (0, 2, 4, or 6 hours) centered at 12:00 PM. Details on experimental interventions are provided below and in the Table.

**LIM Group.** Chicks in this group (n = 13) were raised under background laboratory light conditions and were not subjected to HL or UnV.

**High-Intensity Light Groups (LIM + HL).** Three groups (n = 13 each) were exposed to 2, 4, or 6 hours of HL (15,000 lux) per day and background light for the remainder of the light cycle. Defocusing lenses were not removed during HL.

**Unrestricted Vision Groups (LIM + UnV).** Three groups (n = 13, 12, and 11) were raised under background light during the light cycle and throughout the experiment. Defocusing lenses were removed for 2, 4, or 6 h/d.

High-Intensity Light and Unrestricted Vision Groups (LIM + HL + UnV). Three groups (n = 12, 13, and 13) were exposed to 2, 4, or 6 hours of HL (15,000 lux)

per day and background light for the remainder of the light cycle. Defocusing lenses were removed during HL.

#### **Ocular Measurements In Vivo**

On D1, day 4 (D4), and D8, body weight, ocular AL, refractive error, choroidal thickness (CT), central corneal thickness (CCT), and anterior chamber depth (ACD) were measured in the experimental and control eyes of alert, gently handheld chicks. Measurements were taken in a dimly lit room (<5 lux) on all chicks, in a random order, between 12 PM and 5 PM to reduce any impact of circadian rhythm on outcome measures. A lid retractor was used in only a small number of chicks (2 to 3 animals per group, especially on D1) who would not keep their eyelids open. Whenever used, the lid retractor was inserted very carefully by the experimenter so that it does not come into contact with the cornea or obstruct the ophthalmic examination procedure.

# **Axial Length**

AL was measured using VuMAX HD (Sonomed Escalon, New Hyde Park, NY, USA) A-scan ultrasonography with a probe frequency of 10 MHz as described elsewhere.<sup>41</sup> AL was measured as the distance between the echo spike corresponding to the anterior surface of the cornea and most anterior spike originating from the retina. Each recorded measurement was the median of 7 to 10 scans.

# Refraction

Ocular refraction was measured using a calibrated automated infrared photo-retinoscope as previously described.<sup>42</sup> Alert chicks were handheld on an adjustable platform ~1 m away from the infrared photo-retinoscope. The chick's head was carefully positioned to ensure optimal focus on the chick's eye and first Purkinje image. Pupil size was adjusted for each eye. The median of the most hyperopic refraction readings (i.e., resting refraction) with no accommodative changes was calculated from the continuous refraction trace comprising at least 300 readings over time in each eye.<sup>28,42</sup>

# Choroidal Thickness and Anterior Segment Characteristics

CT at the posterior pole was measured using posterior segment optical coherence tomography (OCT) (Spectralis;

Heidelberg Engineering, Inc., Heidelberg, Germany), and anterior segment (CCT and ACD) was imaged using anterior segment OCT (RTVue; Optovue, Inc., Fremont, CA, USA) as per the protocols described in Najjar et al.<sup>41</sup> During both procedures, the alert chick's head was gently held and aligned with the OCT camera lens such that the infrared laser beam entered the eve through the center of the pupil. The OCT operator further refined the centration of the pupil, and multiple OCT scans were captured. For posterior segment OCT measurement, the centration was within  $\pm 100 \ \mu m$  from the horizontal line. Distance between the inner border of the sclera and the outer border of the retinal pigment epithelium (RPE) was defined as the CT. The average of three thickness measurements of the central cornea was defined as the CCT, whereas ACD was defined as the distance between the central most posterior layer of the cornea and the central most anterior layer of the lens. Measurements were done manually by the first author (SB), who, during measurement sessions, was kept blinded to the eye (LIM or control) and study group (HL, UnV, HL + UnV) conditions.

# **Analyses and Statistics**

All data are represented as mean  $\pm$  SEM of the interocular difference (IOD) between the experimental and uncovered control eyes (i.e., experimental [LIM] eye - control eye) to account for interanimal differences in outcome measures, given the mixed breed and large number of animals (n = 126 chicks) used in this study. Two-way repeatedmeasures ANOVA with the factors day, group, and the group × day interaction was used to compare IODs in refraction, AL, CT, ACD, and CCT. A Holm-Sidak method for pairwise multiple comparisons was performed when the omnibus test for interaction effects between group  $\times$  day was significant. A two-way ANOVA was used to evaluate the interaction between the type of intervention (HL, UnV, HL + UnV) and its duration (0, 2, 4, and 6 hours) on refraction, AL, and CT. A Holm-Sidak method for pairwise multiple comparisons was used when the omnibus test reached statistical significance. The significance level for all statistical tests was set at  $\alpha = 0.05$  with Sidak correction for post hoc pairwise comparisons.

#### RESULTS

#### **Ocular Changes Associated With LIM**

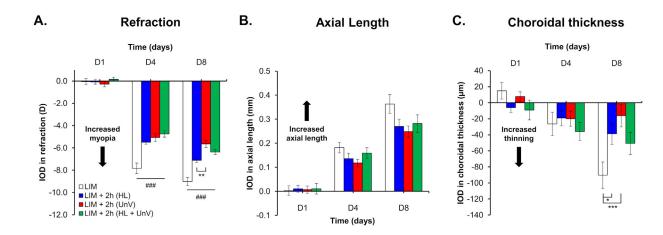
Compared to the uncovered contralateral control eyes (refraction: +4.83 ± 0.05 D and +5.27 ± 0.06 D by D4 and D8, respectively), the LIM eyes (refraction:  $-3.00 \pm 0.11$  D and  $-3.75 \pm 0.07$  D by D4 and D8, respectively) displayed a myopic shift in refractive error that predominantly occurred within the first 4 days after -10 D lens wear (IOD:  $-7.83 \pm 0.48$  D and  $-9.02 \pm 0.37$  D by D4 and D8, respectively). Concurrently, LIM eyes showed an increase in AL (IOD:  $+0.18 \pm 0.02$  mm and  $+0.36 \pm 0.04$  mm by D4 and D8, respectively) and a decrease in CT (IOD:  $-26.35 \pm 14.55 \mu$ m and  $-90.27 \pm 16.44 \mu$ m by D4 and D8, respectively) compared to control eyes (all, *P* < 0.001) (Figs. 2, 3, and 4, Supplementary Table S1). The CCT and ACD of LIM eyes were not different from control eyes.

#### Impact of 2 Hours of HL, UnV, and HL + UnV

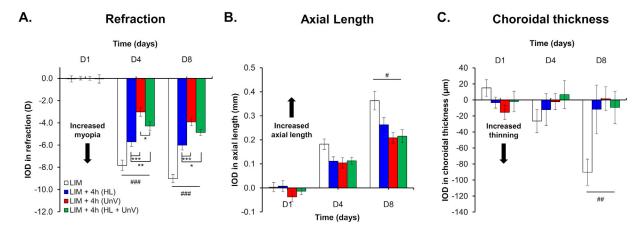
For 2-hour interventions, there was a significant interaction between group and day of intervention for IOD in refraction (F(6, 94) = 10.29, P < 0.001). By D4 and D8, 2 hours of HL, UnV, and HL + UnV significantly reduced myopic refraction compared to the LIM group (both for D4 and D8, P < 0.001). Two hours of UnV, however, was more efficient than 2 hours of HL in reducing myopic refraction on D8 (P = 0.001). HL + UnV was as effective as HL and UnV in reducing myopic refraction on D4 and D8 (P > 0.05) (Fig. 2A). IOD in AL was only dependent on the day of the intervention (F(2,47) = 161.3, P < 0.001). Differences between groups did not reach statistical significance (Fig. 2B). The group  $\times$  day interaction for IOD in CT was significant (F(6, 94) = 3.61), P = 0.003) with UnV (P < 0.001) and HL (P = 0.01) significantly reducing choroidal thinning on D8 but not on D4 (Fig. 2C). The detailed results are provided in Supplementary Table S1.

#### Impact of 4 Hours of HL, UnV, and HL + UnV

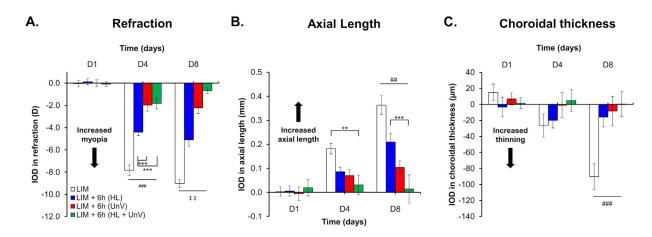
For 4-hour interventions, we found a significant group  $\times$  day interaction for IOD in refraction (*F*(6, 94) = 12.80,



**FIGURE 2.** IOD in refraction, axial length, and choroidal thickness on days 1, 4, and 8 of the experimental protocol in the group not exposed to any intervention (LIM) and groups exposed to 2 hours of HL, UnV, or both (HL + UnV). All groups are significantly different from the LIM group:  $^{###}P < 0.001$ ,  $^{##}P < 0.01$ ,  $^{#P}P < 0.05$ . For significant group effect:  $^{*}P < 0.05$ ,  $^{**}P < 0.01$ ,  $^{***}P < 0.001$  (two-way repeated-measures ANOVA).



**FIGURE 3.** IOD in refraction, axial length, and choroidal thickness on days 1, 4, and 8 of the experimental protocol in the group not exposed to any intervention (LIM) and groups exposed to 4 hours of HL, UnV, or both (HL + UnV). All groups are significantly different from the LIM group:  $^{###}P < 0.001$ ,  $^{##}P < 0.01$ ,  $^{#P}P < 0.05$ . For significant group effect:  $^{*}P < 0.05$ ,  $^{**}P < 0.01$ ,  $^{***}P < 0.001$  (two-way repeated-measures ANOVA).

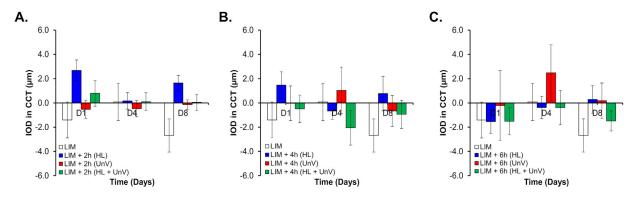


**FIGURE 4.** IOD in refraction, axial length, and choroidal thickness on days 1, 4, and 8 of the experimental protocol in the group not exposed to any intervention (LIM) and groups exposed to 6 hours of HL, UnV, or both (HL + UnV). All groups are significantly different from the LIM group:  ${}^{\#\#}P < 0.001$ ,  ${}^{\#}P < 0.01$ ,  ${}^{\#}P < 0.05$ . All groups are significantly different from each other:  ${}^{\#\#}P < 0.001$ ,  ${}^{\#}P < 0.01$ ,  ${}^{\#}P < 0.05$ . For significant group effect:  ${}^{*}P < 0.05$ ,  ${}^{*}P < 0.001$  (two-way repeated-measures ANOVA).

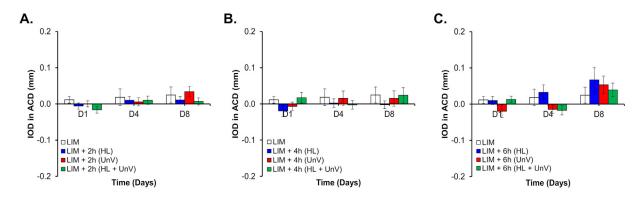
P < 0.001). By D4 and D8, 4 hours of HL, UnV, and HL + UnV significantly reduced myopic refraction compared to the LIM group (both for D4 and D8, P < 0.001). On D4 and D8, 4 hours of UnV was more effective than HL (P < 0.001) in reducing myopic refraction, while HL + UnV was more effective than HL in reducing myopic refraction (D4: P = 0.009, D8: P = 0.04). On D4 but not on D8, UnV was more effective than HL + UnV (P = 0.01) in reducing myopic refraction (Fig. 3A). A significant group  $\times$  day interaction was also found for IOD in AL (F(6, 94) = 2.23, P = 0.047), with all groups showing significantly reduced axial elongation compared to that observed in the LIM group (D4: no difference between groups; D8: LIM versus UnV: P < 0.001; LIM versus HL: P = 0.01; LIM versus HL + UnV: P < 0.001) (Fig. 3B). Differences in AL between intervention groups did not reach statistical significance. The group  $\times$  day interaction for IOD in CT was significant (F(6, 94) = 3.98), P = 0.001). On D4, differences in CT between groups did not reach statistical significance. On D8, all intervention groups prevented choroidal thinning in the myopic eye compared to the LIM group on D8 (LIM versus UnV: P < 0.001; LIM versus HL: P = 0.002; LIM versus HL + UnV: P = 0.002) (Fig. 3C). The detailed results are provided in Supplementary Table S1.

#### Impact of 6 Hours of HL, UnV, and HL + UnV

Six-hour interventions had a significant interaction between the group and time for IOD in refraction (*F*(6, 92) = 24.67, *P* < 0.001). By D4 and D8, 6 hours of HL, UnV, and HL + UnV significantly reduced myopic refraction compared to the LIM group (both D4 and D8, *P* < 0.001). Six hours of UnV was more effective than HL in reducing myopic refraction by both D4 and D8 (*P* < 0.001). HL + UnV was more effective in reducing myopic refraction than both HL (both D4 and D8: *P* < 0.001) and UnV (only D8: *P* = 0.008) (Fig. 4A). IOD in AL had a significant group × time interaction (*F*(6, 92) = 6.4, *P* < 0.001) with all groups showing reduced axial elongation compared to the LIM eyes (LIM versus UnV: *P* < 0.001; LIM versus HL: *P* = 0.004; LIM versus HL + UnV: *P* < 0.001) on D8. On D4, only the HL + UnV group showed



**FIGURE 5.** IOD in CCT on days 1, 4, and 8 of the experimental protocol in the group not exposed to any intervention (LIM) and groups exposed to 2 hours (**A**), 4 hours (**B**), and 6 hours (**C**) of HL, UnV, or both (HL + UnV).

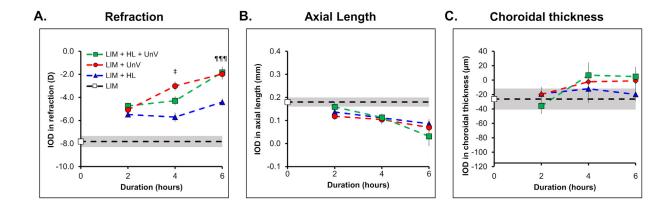


**FIGURE 6.** IOD in ACD on days 1, 4, and 8 of the experimental protocol in the group not exposed to any intervention (LIM) and groups exposed to 2 hours (**A**), 4 hours (**B**), and 6 hours (**C**) of HL, UnV, or both (HL + UnV).

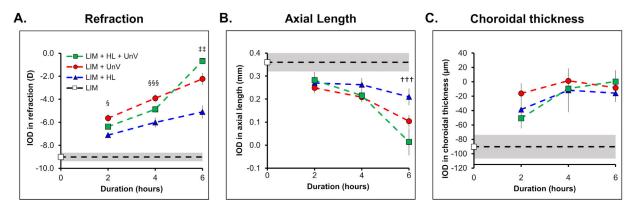
significantly reduced axial elongation compared with the LIM group (P = 0.009). HL + UnV was more effective than HL in reducing axial elongation (P < 0.001) on D8 but not on D4 (Fig. 4B). CT also showed a significant interaction between groups and time (F(6, 92) = 3.53, P = 0.003), with all three interventions significantly reducing choroidal thinning of the myopic eye compared to the LIM group on D8 (LIM versus all groups: P < 0.001) (Fig. 4C). Differences in CT between intervention groups did not reach statistical significance.

# Impact of Experimental Interventions on ACD and CCT

IODs in CCT were only significantly different (F(3, 94) = 3.86, P = 0.02) between the HL and LIM groups (P = 0.01) for 2-hour-long interventions. IODs in ACD showed a significant effect of day only for the 2-hour (F(2, 94) = 3.99, P = 0.02) and 6-hour (F(2, 92) = 7.31, P = 0.001) interventions (Figs. 5 and 6). For more details, see Supplementary Table S1.



**FIGURE 7.** Duration-response curve for the IOD in refraction (**A**), axial length (**B**), and choroidal thickness (**C**) in the groups exposed to 2, 4, and 6 hours of HL, UnV, or both (HL + UnV) on day 4 of the experimental protocol. The LIM group that was not exposed to any intervention is represented by a *white square* and a *shaded area* for mean  $\pm$  95% confidence interval. All groups are different from each other:  ${}^{\ddagger}P < 0.05$ . HL significantly different from both UnV and HL + UnV:  ${}^{\$\$}P < 0.001$ .



**FIGURE 8.** Duration-response curve for the IOD in refraction (**A**), axial length (**B**), and choroidal thickness (**C**) in the groups exposed to 2, 4, and 6 hours of HL, UnV, or both (HL + UnV) on day 8 of the experimental protocol. The LIM group that was not exposed to any intervention is represented by a *white square* and a *shaded area* for mean  $\pm$  95% confidence interval. HL is significantly different from UnV: <sup>§</sup>*P* < 0.05, <sup>§§§</sup>*P* < 0.001. All groups are different from each other (at least, <sup>‡‡</sup>*P* < 0.01). HL is significantly different from HL + UnV (<sup>††</sup>*P* < 0.001).

# Duration Response Curves on D4 and D8 of the Interventions

On D4 of the protocol, the impact of the intervention on IODs of refraction (F(3, 140) = 80.93, P < 0.001) and AL (F(3, 140) = 14.49, P < 0.001) was duration dependent. The interaction between the group and time for IOD in refraction was significant (F(6, 140) = 3.63, P = 0.002), where both 4 and 6 hours of UnV (both 4 and 6 hours: P < 0.001) and HL + UnV (4 hours: P = 0.03 and 6 hours: P < 0.001) were more effective than HL in reducing myopic refraction. UnV of 4 hours but not 6 hours reduced myopic refraction more than HL + UnV (P = 0.03) (Fig. 7A). IODs in AL and CT were not different between groups across the different durations of the interventions (Figs. 7B, 7C).

On D8 of the protocol, the impact of interventions (i.e., HL, UnV, HL + UnV) on IODs of refraction (F(2, 104) = 78.33,P < 0.001) and AL (F(2, 104) = 17.1, P < 0.001) was duration dependent. There was a significant interaction between the duration and type of intervention on IODs of refraction (F(4,104) = 8.38, P < 0.001), where only 6-hour HL + UnV but not 2 or 4 hours of HL + UnV, was more effective than UnV (P = 0.004) and HL (P < 0.001) in reducing myopic refraction. Conversely, regardless of the duration, UnV was more effective than HL in reducing myopic refraction (2 hours: P = 0.01; 4 and 6 hours: P < 0.001) (Fig. 8A). Likewise, the interaction between the duration and type of intervention was significant for AL (F(4, 104) = 2.60, P = 0.04), where only 6 hours of HL + UnV was more effective than HL (P < 0.001) (Fig. 8B). IOD of CT was not different between intervention groups across the different durations (Fig. 8C).

# DISCUSSION

In this study, we demonstrated that different combinations and durations of HL and UnV have varying effects on the development of LIM in a chicken model. The effect of HL alone, UnV alone, and HL + UnV in reducing myopia and AL elongation was duration dependent (IODs for refraction and AL were lowest at 6 hours, followed by 4 hours, then 2 hours). We also observed that UnV alone was more effective than HL alone in preventing LIM. Interestingly, the effect of HL + UnV in reducing LIM development was only found to be additive to a statistically significant extent (lower IODs

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than both UnV alone and HL alone) in the 6-hour exposure group. On D8, the greatest reduction in LIM (closest to null IODs for myopia, AL, and CT) was found in chicks subjected to 6-hour daily exposure of HL + UnV. We did not find any significant change in ACD or CCT among the groups, which was consistent with previous findings.<sup>30</sup>

The duration-dependent effect of UnV in reducing LIM development has been previously reported in guinea pigs,<sup>43</sup> chickens,44 tree shrews,33 and marmosets.34 In chickens, using younger (D1-D10) and older (D7-D11) animals, Schmid and Wildsoet<sup>44</sup> reported a ~90% reduction in LIM (-10 D) by just 3 hours of normal vision or UnV per day. Contrarily, in older chicks (D7-D11) with no accommodation due to ciliary nerve section, 3 and 6 hours of UnV reduced their LIM by 60% and 96%, respectively, beyond which (i.e., 9-hour UnV) the effect plateaued. In comparison, we observed 37%, 57%, and 75% reduction in LIM by D8 on exposure to 2, 4, and 6 hours of UnV, respectively. The reduced impact of UnV that we report in our study may be due to experimental protocol differences, particularly differences in the age (visual maturation) and strain of chickens, the timing of UnV (i.e., centered at noon), experimental protocol duration, and the characteristics of the background lighting and visual-spatial environments during normal vision/UnV. In addition, Nickla et al.<sup>45</sup> showed less ocular growth and LIM during the afternoon than in the morning in response to -10 D defocus in chicks. The authors also found UnV to be more effective to reduce LIM in the evening than in the morning.<sup>46</sup> From our findings, it can be argued that, when centered at noon, longer durations of continuous UnV, spilling further into the afternoon (i.e., 2 hours: 11 AM to 1 PM; 4 hours: 10 AM to 2 PM; 6 hours: 9 AM to 3 PM), would potentiate the impact of UnV further. Conversely, here we show a newfound duration-dependent impact of HL on reducing myopic shift and axial elongation in LIM. There is currently insufficient evidence linking the total quantum of light exposure (intensity of the exposure  $\times$  daily duration of the exposure  $\times$  duration of the study protocol) with LIM reduction, but a recent study investigating LIM in mice showed that retinal expression of DA-related genes and proteins increased with increasing intensities of light,<sup>38</sup> suggesting a dose-dependent release of ocular growth neuromodulators like DOPAC, a metabolite of DA, and/or nitric oxide (NO).38,47,48 Whether this release

is dependent on the total quantum of light, whereby the retina acts as a photon counter to drive neuromodulatory responses, remains unclear yet unlikely, given that short intermittent light pulses have been reported to be more effective than equiluminant continuous light in chickens.<sup>49</sup> In humans, myopia progression rates in children are slower during a 6-month period that included summer vacation, when days are longer and brighter, compared to winter. These findings partially support an association between the total quantum of light exposure and myopia control.<sup>50</sup> On the other hand, it is worth mentioning that Stone and colleagues<sup>51</sup> reported only a partial and transient effect (i.e., on D4 but not on D11 of the authors' experimental protocol) of natural high-intensity daylight on FDM in chicks reared outdoors, without any established associations with the retinal DA levels. Given that our experimental protocol was only 8 days long, a longer investigation with more frequent data sampling is required to confirm the permanency of the duration-dependent impact of HL on LIM.

Next, UnV alone was more effective than HL alone in reducing LIM development. During UnV, a temporary myopic defocus occurs, while in HL, the hyperopic defocus is constantly present in eyes with LIM. Studies had shown that when the eye receives competing defocus signals, the more myopic defocus dominates ocular growth control<sup>52-54</sup>; this could explain the stronger emmetropization signal from UnV. Another suggestion for the disparity involves UnV being a visually/optically guided feedback process,<sup>15,52</sup> with an endpoint of refractive error elimination. In contrast, HL acting through a potentially distinct pathway involves photoreceptor stimulation and retinal neuromodulation<sup>26,29,30</sup> that can only marginally overcome the existing visual experience (i.e., hyperopic defocus). Whether UnV remains a more potent driver for emmetropization if the myopiagenic stimulus is reduced (e.g., using a -5 D defocus instead of -10 D) remains to be investigated. Additionally, axial myopic retinas are more prolate (flatter in periphery) and have a relative peripheral hyperopia compared to hyperopic eyes with oblate retinas (steeper in periphery) and relative peripheral myopia.55,56 Relative peripheral refraction (hyperopia or myopia) compared to the central refraction may exacerbate or reduce myopia development, respectively,55,56 and can partially explain a greater drive of UnV alone in reducing LIM compared to HL alone.

Our data did not demonstrate that the combined effect of HL + UnV in reducing LIM development was additive, when interventions were 2 or 4 hours long. These findings suggested that UnV and HL may be deploying different pathways of compensatory mechanisms in LIM.<sup>18</sup> In chicks, Ashby et al.<sup>29</sup> found that on day 5, the protective effects of diffuser removal against FDM was significantly enhanced by 15 minutes of high-intensity indoor lights (15,000 lux) during diffuser removal. This was not in agreement with our findings, where at both D4 and D8, the protective impact of HL + UnV on AL increase, myopic refraction, and choroidal thinning was similar if not worse than UnV alone for 2- and 4-hour-long exposures. The additional alleviation from FDM observed by Ashby and colleagues may result from higher exposure levels to HL and DA synthesis due to diffuser removal,<sup>36-38</sup> while in LIM, there was only a minimal increase in HL levels reaching the eye associated with UnV. In addition, Ashby et al.<sup>29</sup> reported that the spectral distribution of their experimental halogen lights (300-1000 nm, peaking at 700 nm) was similar to daylight over the range of visible light for chickens (360-700 nm),<sup>57</sup> while

our experimental light had a typical LED spectrum peaking around 449 nm and 583 nm (Fig. 1). The fullness of the halogen light spectrum light may have promoted the impact of short-duration, high-intensity light on the recovery from FDM.<sup>28</sup> Parenthetically, studies investigating the impact of lens or diffuser removal should take into account the different spatial characteristics of the housing environment.<sup>12,39</sup> Another plausible explanation for our findings was that the significant increase in pupillary constriction induced by HL, during UnV, increased the depth of focus and attenuated the accuracy of the UnV drive. Pupillary constriction under HL may also explain the reduced impact of defocus on LIM but failed to explain the potentiating effect of HL on lensinduced hyperopia.<sup>30</sup>

Interestingly, HL and UnV were additive to a statistically significant extent when interventions were 6 hours long and almost completely compensated for LIM (IOD: -0.68 D), AL (IOD: 0.01 mm), and CT (IOD: 0.46 µm) by D8 (Supplementary Table S1). These findings suggested a durationdependent synergy/intermodulation between HL and UnV. While we do not have an understanding of the mechanisms to explain these data yet, this duration-dependent synergy may originate from delayed photobiomodulatory processes originating at the retinal level that enhance the drive for emmetropization conferred by UnV.58 Vitreal DOPAC (a metabolite of DA) levels are known to increase with both the duration<sup>59</sup> and intensity<sup>37</sup> of light exposure in chickens, with the circadian rhythm of DA release observed to peak at 12 hours into the light phase.<sup>59</sup> DA can trigger the release of other neurotransmitters like NO,47 reducing LIM either alone or in conjunction. This DA release is shown to depend on not only the retinal luminance but also the image contrast/visual spatial information,<sup>60</sup> which gets altered during normal vision or UnV. It is possible that the combination of UnV and HL for 6 hours surpassed a retinal DA threshold that enhanced recovery from LIM further, through an overlapping pathway between UnV and HL. It is worth highlighting that the trend of HL and UnV alone was unaltered throughout the different durations of exposure.

In existing literature, chicks had shown changes in CT in response to altered retinal image quality, and it had been suggested that this CT is a part of mechanisms regulating ocular growth and emmetropization.<sup>2,61,62</sup> As expected, in our study, LIM led to choroidal thinning.45,46 While eyes exposed to 4 and 6 hours of HL, UnV, or HL + UnV all had significantly thicker choroids compared to LIM eyes, 2 hours of HL and UnV but not HL + UnV led to significantly thicker choroids than LIM eyes, corroborating the observed nonadditivity of 2-hour-long HL and UnV exposures for AL and refraction. A reduction in choroidal thinning was particularly observed when the duration of interventions was increased from 2 to 4 hours. Although these findings may be explained by a rise in intraocular temperature, leading to increased permeability and vasodilation of choroidal vessels, and a release of neurotransmitters resulting in choroidal thickening,  $^{61,62}$  it is unlikely, yet untested, that our wide-field HL intervention caused any changes in intraocular temperature. Nonetheless, the synergy between HL and UnV at 6-hour interventions seems to be independent of choroidal thickening induced by HL as 4 hours of HL yielded similar choroidal thickening as 6 hours of HL.

Our study has a few limitations. First, we have opted for an environment lacking color, movement, and other usual features of a visual environment for the housing of the chicks. This environment lacks fine spatial details and may not be the most suitable environment for emmetropization or the recovery from LIM.12 Second, given the differences in ocular optics, anatomy, and physiology between chickens and humans,<sup>63</sup> it was difficult to directly contextualize and translate our findings to humans. While our work is in agreement with the literature that removing hyperopic defocus reduces myopia development, such findings are applicable to animal models with normally functioning emmetropization vet potentially less so to myopic human subjects with deficiencies in the emmetropization mechanism.<sup>64</sup> Finally, our findings suggest that exposure to HL can slow the development of myopia in a duration-dependent manner regardless of the refractive status of the eye or concurrent myopiagenic stimuli (e.g., near visual work). In addition, the combination of high-intensity light exposure and visual breaks (e.g., reducing accommodative lag) may have the highest potential to prevent myopia development, when applied for long durations during daytime (50% of daytime here). These lengthy interventions may not be implementable in real life.

# **CONCLUSION**

In this study, we showed that daily exposure to 2, 4, or 6 hours of HL or UnV slows the shift to myopic refraction in chickens in a duration-dependent manner. Combined with UnV, only 6 hours of HL fully halted the development of LIM (i.e., axial elongation, myopic refraction, and choroidal thinning). The synergetic effect of HL and UnV is dependent on the duration of the intervention. Further molecular work is required to better understand this peculiar synergy between HL and UnV, with the potential of translating such findings into pharmacologic interventions or combined light/optics interventions.

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#### References

- 1. Morgan IG, French AN, Ashby RS, et al. The epidemics of myopia: Aetiology and prevention. *Prog Retin Eye Res.* 2018;62:134–149.
- 2. Baird PN, Saw SM, Lanca C, et al. Myopia. Nat Rev Dis Primers. 2020;6(1):1-20.
- 3. Bullimore MA, Brennan NA. Myopia control: Why each diopter matters. *Optom Vis Sci.* 2019;96(6):463–465.
- Morgan IG, Wu PC, Ostrin LA, et al. IMI risk factors for myopia. *Invest Ophthalmol Vis Sci.* 2021;62(5):3.
- 5. Jin JX, Hua WJ, Jiang X, et al. Effect of outdoor activity on myopia onset and progression in school-aged children in

northeast China: The Sujiatun Eye Care Study. *BMC Ophthalmol.* 2015;15(1):1–11.

- 6. Wu PC, Chen CT, Lin KK, et al. Myopia prevention and outdoor light intensity in a school-based cluster randomized trial. *Ophthalmology*. 2018;125(8):1239–1250.
- 7. He M, Xiang F, Zeng Y, et al. Effect of time spent outdoors at school on the development of myopia among children in China: A randomized clinical trial. *JAMA*. 2015;314(11):1142–1148.
- 8. Guggenheim JA, Northstone K, McMahon G, et al. Time outdoors and physical activity as predictors of incident myopia in childhood: A prospective cohort study. *Invest Ophthalmol Vis Sci.* 2012;53(6):2856–2865.
- McKnight CM, Sherwin JC, Yazar S, et al. Myopia in young adults is inversely related to an objective marker of ocular sun exposure: The Western Australian Raine cohort study. *Am J Ophthalmol.* 2014;158(5):1079–1085.
- 10. Najjar RP. Opinion: Tailoring the lighting environment for a healthier ocular growth. *Lighting Res Technol*. 2022;54(2):100.
- 11. Flitcroft DI. The complex interactions of retinal, optical and environmental factors in myopia aetiology. *Prog Retin Eye Res.* 2012;31(6):622–660.
- 12. Flitcroft DI, Harb EN, Wildsoet CF. The spatial frequency content of urban and indoor environments as a potential risk factor for myopia development. *Invest Ophtbalmol Vis Sci.* 2020;61(11):42.
- 13. Ngo C, Saw SM, Dharani R, Flitcroft I. Does sunlight (bright lights) explain the protective effects of outdoor activity against myopia? *Ophthalmic Physiol Optics*. 2013;33(3):368– 372.
- Muralidharan AR, Lança C, Biswas S, et al. Light and myopia: From epidemiological studies to neurobiological mechanisms. *Ther Adv Ophthalmol.* 2021;13(D):251584142110592.
- 15. Carr BJ, Stell WK. The science behind myopia. In: Kolb H, Fernandez E, Nelson R, eds. Webvision: The Organization of the Retina and Visual System [Internet], Salt Lake City (UT): University of Utah Health Sciences Center; 2017, http://www.ncbi.nlm.nih.gov/pubmed/29266913. Accessed February 18, 2023.
- Morgan IG, Ashby RS, Nickla DL. Form deprivation and lensinduced myopia: Are they different? *Ophthalmic Physiol Opt*. 2013;33(3):355–361.
- 17. Nickla DL, Totonelly K. Dopamine antagonists and brief vision distinguish lens-induced- and form-deprivation-induced myopia. *Exp Eye Res.* 2011;93(5):782–785.
- Nickla DL, Yusupova Y, Totonelly K. The muscarinic antagonist MT3 distinguishes between form deprivation-and negative lens-induced myopia in chicks. *Curr Eye Res.* 2015;40(9):962–967.
- 19. Mertz JR, Wallman J. Choroidal retinoic acid synthesis: A possible mediator between refractive error and compensatory eye growth. *Exp Eye Res.* 2000;70(4):519–527.
- 20. Thomson K, Karouta C, Ashby R. Form-deprivation and lensinduced myopia are similarly affected by pharmacological manipulation of the dopaminergic system in chicks. *Invest Ophthalmol Vis Sci.* 2020;61(12):4.
- 21. Thomson K, Morgan I, Karouta C, Ashby R. Levodopa inhibits the development of lens-induced myopia in chicks. *Sci Rep.* 2020;10(1):1–12.
- 22. Ashby R, Kozulin P, Megaw PL, Morgan IG. Alterations in ZENK and glucagon RNA transcript expression during increased ocular growth in chickens. *Mol Vis.* 2010;16:639.
- 23. Xiao H, Fan ZY, Tian XD, Xu YC. Comparison of formdeprived myopia and lens-induced myopia in guinea pigs. *Int J Ophthalmol.* 2014;7(2):245.
- 24. Schmid KL, Wildsoet CF. Inhibitory effects of apomorphine and atropine and their combination on myopia in chicks. *Optom Vis Sci.* 2004;81(2):137–147.

- 25. Zhang L, Qu X. The effects of high lighting on the development of form-deprivation myopia in guinea pigs. *Invest Ophthalmol Vis Sci.* 2019;60(13):4319–4327.
- 26. Karouta C, Ashby RS. Correlation between light levels and the development of deprivation myopia. *Invest Ophthalmol Vis Sci.* 2015;56(1):299–309.
- 27. Smith EL, Hung LF, Huang J. Protective effects of high ambient lighting on the development of form-deprivation myopia in rhesus monkeys. *Invest Ophthalmol Vis Sci.* 2012;53(1):421–428.
- 28. Muralidharan AR, Low SWY, Lee YC, et al. Recovery from form-deprivation myopia in chicks is dependent upon the fullness and correlated color temperature of the light spectrum. *Invest Ophthalmol Vis Sci.* 2022;63(2):16.
- 29. Ashby R, Ohlendorf A, Schaeffel F. The effect of ambient illuminance on the development of deprivation myopia in chicks. *Invest Ophthalmol Vis Sci.* 2009;50(11):5348– 5354.
- Ashby RS, Schaeffel F. The effect of bright light on lens compensation in chicks. *Invest Ophthalmol Vis Sci.* 2010;51(10):5247–5253.
- 31. Smith EL, Hung LF, Arumugam B, Huang J. Negative lensinduced myopia in infant monkeys: Effects of high ambient lighting. *Invest Ophthalmol Vis Sci.* 2013;54(4):2959–2969.
- 32. McBrien NA, Arumugam B, Metlapally S. The effect of daily transient +4 D positive lens wear on the inhibition of myopia in the tree shrew. *Invest Ophthalmol Vis Sci.* 2012;53(3):1593–1601.
- 33. Shaikh AW, Siegwart JTJ, Norton TT. Effect of interrupted lens wear on compensation for a minus lens in tree shrews. *Optom Vis Sci.* 1999;76(5):308–315.
- 34. Benavente-Perez A, Nour A, Troilo D. Short interruptions of imposed hyperopic defocus earlier in treatment are more effective at preventing myopia development. *Sci Rep.* 2019;9(1):25–29.
- 35. Nickla DL. Transient increases in choroidal thickness are consistently associated with brief daily visual stimuli that inhibit ocular growth in chicks. *Exp Eye Res.* 2007;84(5):951–959.
- Brainard GC, Morgan WW. Light-induced stimulation of retinal dopamine: A dose-response relationship. *Brain Res.* 1987;424(1):199–203.
- 37. Cohen Y, Peleg E, Belkin M, Polat U, Solomon AS. Ambient illuminance, retinal dopamine release and refractive development in chicks. *Exp Eye Res.* 2012;103:33–40.
- Landis EG, Park HN, Chrenek M, et al. Ambient light regulates retinal dopamine signaling and myopia susceptibility. *Invest Ophthalmol Vis Sci.* 2021;62(1):28.
- Schmid KL, Wildsoet CF. Contrast and spatial-frequency requirements for emmetropization in chicks. Vis Res. 1997;37(15):2011–2021.
- Schippert R, Schaeffel F. Peripheral defocus does not necessarily affect central refractive development. *Vis Res.* 2006;46(22):3935–3940.
- 41. Najjar R, Chao De La, Barca JM, Barathi VA, et al. Ocular growth and metabolomics are dependent upon the spectral content of white light. *Invest Ophthalmol Vis Sci.* 2018;59(9):755.
- Schaeffel F, Farkas L, Howland HC. Infrared photoretinoscope. *Appl Opt.* 1987;26(8):1505.
- 43. Leotta AJ, Bowrey HE, Zeng G, Mcfadden SA. Temporal properties of the myopic response to defocus in the guinea pig. *Ophtbalmic Physiol Opt.* 2013;33(3):227–244.
- 44. Schmid KL, Wildsoet CF. Effects on the compensatory responses to positive and negative lenses of intermittent lens wear and ciliary nerve section in chicks. *Vision Res.* 1996;36(7):1023–1036.

- 45. Nickla DL, Jordan K, Yang J, Totonelly K. Brief hyperopic defocus or form deprivation have varying effects on eye growth and ocular rhythms depending on the time-of-day of exposure. *Exp Eye Res.* 2017;161:132–142.
- 46. Nickla DL, Thai P, Zanzerkia Trahan R, Totonelly K. Myopic defocus in the evening is more effective at inhibiting eye growth than defocus in the morning: Effects on rhythms in axial length and choroid thickness in chicks. *Exp Eye Res.* 2017;154:104–115.
- 47. Zhou X, Pardue MT, Iuvone PM, Qu J. Dopamine signaling and myopia development: What are the key challenges. *Prog Retin Eye Res.* 2017;61:60–71.
- 48. Sekaran S, Cunningham J, Neal MJ, Hartell NA, Djamgoz MB. Nitric oxide release is induced by dopamine during illumination of the carp retina: Serial neurochemical control of light adaptation. *Eur J Neurosci.* 2005;21(8):2199–2208.
- 49. Lan W, Feldkaemper M, Schaeffel F. Intermittent episodes of bright light suppress myopia in the chicken more than continuous bright light. *PLoS One.* 2014;9(10):1–7.
- Gwiazda J, Deng L, Manny R, Norton TT, for the COMET Study Group. Seasonal variations in the progression of myopia in children enrolled in the correction of myopia evaluation trial. *Invest Ophthalmol Vis Sci.* 2014;55(2):752– 758.
- 51. Stone RA, Cohen Y, McGlinn AM, et al. Development of experimental myopia in chicks in a natural environment. *Invest Ophthalmol Vis Sci.* 2016;57(11):4779–4789.
- 52. Diether S, Wildsoet CF. Stimulus requirements for the decoding of myopic and hyperopic defocus under single and competing defocus conditions in the chicken. *Invest Ophthalmol Vis Sci.* 2005;46(7):2242–2252.
- 53. Arumugam B, Hung LF, ho To C, Holden B, Smith EL. The effects of simultaneous dual focus lenses on refractive development in infant monkeys. *Invest Ophthalmol Vis Sci.* 2014;55(11):7423–7432.
- 54. Dennis YT, To C. Graded competing regional myopic and hyperopic defocus produce summated emmetropization set points in chick. *Invest Ophthalmol Vis Sci.* 2011;52(11):8056–8062.
- 55. Smith EL. Optical treatment strategies to slow myopia progression: Effects of the visual extent of the optical treatment zone. *Exp Eye Res.* 2013;114:77–88.
- 56. Matsumura S, Kuo AN, Saw SM. An update of eye shape and myopia. *Eye Contact Lens*. 2019;45(5):279–285.
- Wortel JF, Rugenbrink H, Nuboer JFW. The photopic spectral sensitivity of the dorsal and ventral retinae of the chicken. J Comp Physiol A. 1987;160(2):151–154.
- Zhang P, Zhu H. Light signaling and myopia development: A review. *Ophtbalmol Ther*. 2022;11(3):939–957.
- 59. Megaw PL, Boelen MG, Morgan IG, Boelen MK. Diurnal patterns of dopamine release in chicken retina. *Neurochem Int.* 2006;48(1):17–23.
- Feldkaemper M, Diether S, Kleine G, Schaeffel F. Interactions of spatial and luminance information in the retina of chickens during myopia development. *Exp Eye Res.* 1999;68(1):105–115.
- 61. Shih YF, Fitzgerald ME, Cuthbertson SL, Reiner A. Influence of ophthalmic nerve fibers on choroidal blood flow and myopic eye growth in chicks. *Exp Eye Res.* 1999;69(1):9–20.
- 62. Fitzgerald MEC, Wildsoet CF, Reiner A. Temporal relationship of choroidal blood flow and thickness changes during recovery from form deprivation myopia in chicks. *Exp Eye Res.* 2002;74(5):561–570.
- 63. Schaeffel F, Howland HC, Farkas L. Natural accommodation in the growing chicken. *Vis Res.* 1986;26(12):1977–1993.
- 64. Swiatczak B, Schaeffel F. Myopia: Why the retina stops inhibiting eye growth. *Sci Rep.* 2022;12(1):21704.